

## REEF FISH ASSESSMENT IN FIJI'S "I QOLIQOLI"

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### ABSTRACT

The main objective of the study was to provide an overview of finfish stocks in fishing grounds (*I Qoliqoli*) around Fiji. Under water census was conducted in 16 fishing grounds in different areas of the Fiji Group. Two fishing grounds from each area were surveyed. The abundance and biomass of finfishes in a fishing ground was estimated. The fishing grounds in outer islands have a higher biomass than the main islands (Viti Levu) of the Fiji Group. Abundance and biomass of finfishes among habitats within a fishing ground was also determined. Abundance was higher in the lagoon but somewhat similar in the reef area, reef slope, back reef and fore reef. However, fore reef had higher biomass. There was a slight variation in the size distribution among depths and habitats. Due to the lack of catch data for each fishing ground surveyed it was not possible to determine the status of exploitation of the stock. However, this study implies that highly commercial fishes are rare in areas with higher human population. If more information on catches will be gathered, the data sampled here will improve the data analysis and sampling scheme of the department as well as enhance the gathering of catch data/CPUE during the visual transect surveys in fishing communities.

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## 1. INTRODUCTION

### 1.1. Location

Fiji is located at the heart of the Pacific Ocean, midway between the Equator and the South Pole and lies on longitudes 175°E and latitudes 18°S (Figure 1). Fiji comprises about 330 islands and islets, of which approximately 100 are inhabited. The total land area is approximately 18,300km<sup>2</sup>. The two main islands are Viti Levu and Vanua Levu. Fiji's Exclusive Economic Zone (EEZ) has an area of 1.3 million km<sup>2</sup>. The population in September 2007 was approximately 837,270, of which 57% are indigenous Fijian and most of them live on the coast (Fiji Bureau of Statistics 2009).



Figure 1: Map of Fiji Islands - with approximately 6,437 km<sup>2</sup> of reef (National Geographic Magazine 2004).

### 1.2. Fisheries in Fiji

For centuries, Fiji has relied on the sea and its bountiful resources as a source of food and economic development. In addition, fisheries resources are of cultural significance to the Fijian people (Veitayaki 1995).

The fisheries sector is currently the third largest natural resource sector in Fiji, contributing F\$92 million to GDP or 2.5% of total GDP (Teh *et al.* 2009). The country's fisheries can be placed into six categories which are: coastal commercial (artisanal fishery), coastal subsistence, offshore locally based, offshore foreign based, freshwater and aquaculture (FAO 2009). Offshore fisheries are tuna long lining oriented while coastal or inshore fisheries are for subsistence purposes and artisanal, for sale in local markets and for export (finfish, beachdemer, trochus for buttons, aquarium fish and live rock).

### 1.3. Inshore Fisheries – importance, status and management

The inshore areas are the main domain of subsistence and artisanal fishers (small scale fisheries) (Teh *et al.* 2009). Over 100 species of finfish and 50 species of invertebrates from inshore areas are included in Fiji's fish market statistics (FAO 2009). Artisanal and subsistence fisheries make up 60 – 80% of the nation's catches (Teh *et al.* 2009), emphasizing the importance of inshore fisheries to food security and the economy. Also Starkhouse (2009) estimated that the artisanal and subsistence fisheries, together, deliver an annual catch of over 17,000 tonnes of reef-associated finfish, invertebrates and marine plants, which have a gross value of approximately US\$ 54 million per year. However, the true volume of subsistence catches is uncertain (Teh *et al.* 2009) because landing occurs at every coastal village throughout the country, roughly in proportion to the distribution of the population (FAO 2009), with inadequate financial resources (funds to employ data collectors) to collect subsistence landing data.

About 50 % of all rural households are involved in subsistence fisheries. Starkhouse (2009) estimated that there are more than 28,000 fishers (with and without a fishing license) that rely on Fiji's coral reefs for food and/or income. The per capita consumption of seafood is 44 kg per year, higher than the world average (Asian Developmental Bank 2005). However, it varies between different areas of Fiji.

The common fishing methods used by both artisanal and subsistence fishers are hand line, gleaning (for invertebrates), use of spear/spear gun to dive at night as well as daytime and gillnets. Other methods of fishing include crab trap, push nets, cast nets, line trawling (*vakasavuba*), long spears (*cocoka*), jigging, and fish traps. Derris root (*duva*) is illegal under the fisheries act; however some rural fishers occasionally use it. Nowadays, it is common among artisanal fishermen to dive with a spear gun at night, whereby the catch is greater and larger fish are caught compared to other methods used.

Artisanal fishers more often use boats than subsistence fishers. To fish within the inshore area, fishers used bamboo raft, wooden boats (14 ft) without engine. Fibreglass boats (mostly 23 ft with 40-60 hp) are used by artisanal fishers to travel to long distance fishing ground as well as quickly transporting the catch to major outlets. For the last few years, the department has been assisting artisanal fishermen, providing 23 ft fibreglass boats and fishing gears for tuna fishing around Fishing Aggregating Devices (FAD) under the small scale subsidy scheme (2/3 paid by a fisherman and 1/3 by the government). The intention is to promote offshore fishing relieving pressure on inshore fisheries.

The main fisheries resource use practice in Fiji is the ownership of fishing rights to the customary fishing areas extending to the outer reef slope (Veitayaki 1999). Like the land (Veitayaki 1999), the laws of Fiji recognise the customary rights to fish in traditional fishing grounds (*I Qoliqoli*) and allow the owners of customary rights to dictate the issue of fishing licenses within delineated customary areas by the Fisheries Division (Hunt 1999). Consequently, there are 410 traditional demarcated fishing grounds (*I Qoliqoli*) (Batibasaga 2006) of which people are expected to use their own allocations, and those seeking to use grounds belonging to others are expected to get permission from the owners (Veitayaki 1999).

Artisanal fishers who wish to fish in specific *I Qoliqoli* must obtain fishing licenses from the Fisheries Department, which are renewed every year. However, the village chiefs' consent is

required first before the Fisheries Department issues the license. As of September 2009, the average licensed fishermen recorded by the Fisheries Department are about 5000.

Fiji's inshore fisheries are dynamic with significant changes in resources use each year (Asian Developmental Bank, 2005). Commercial exploitation due to economic demands to which people are subjected and their increased capacity and productivity levels has given most communities the capability to deplete coastal resources rapidly (Veitayaki 1995). As a result, around 70 *I Qoliqoli* are considered to be over exploited, while 250 are considered fully developed (Teh *et al.* 2009). Burke *et al.* (2004) stated that overfishing could be a major pressure on coral reef systems reducing levels of biodiversity and typically resulting in shifts in fish size, abundance, and species composition, altering the ecological balance on the reef. Other major causes of degradation to the coral reef area in Fiji are agricultural activities, beach and sand mining in some areas, building of jetties and groynes (concrete structure that run perpendicular to the shoreline to create beaches), sewage pollution, inappropriate solid waste disposal, soil erosion and siltation, improper disposal of industrial waste in urban areas and natural hazards such as climate change, sea level rise and hurricanes (Aldersberg 2004).

Numerous non-governmental organisations such as World Wild Fund for Nature (WWF), Conservation International (CI), Wildlife Conservation Society (WCS), and Coral Reef InitiativeS for Pacific (CRISP) with the two local institutions, University of the South Pacific (USP) and Fiji Institute of Technology (FIT) are engaged in conservation and inshore sustainable management activities in some areas of Fiji. Department of Fisheries (DOF) is acting as a secretariat of these conservation activities under the Fiji Locally Managed Marine Area (FLMMA) network.

### 1.3.1. Reef finfish in Fiji – importance and status

Fish is an important constituent in the diets in Fiji, as is common throughout the Pacific Islands (Veitayaki, 1995). It is highly desirable part of the diets because of its taste and cultural significance. Fish contribute for almost 25% of the overall protein supply eaten by locals.

The commonly targeted food fish are Lethrinidae (Emperors), Lutjanidae (Snappers), Serranidae (Groupers), Carangidae (Trevallies), Haemulidae (Harlequin sweetlips), Mugilidae (Mulletts), Scombridae (Tunas), Acanthuridae (Surgeonfishes), Scaridae (Parrotfishes), Labridae (Wrasses), Siganidae (Rabbitfishes) and Sphyrinae (Barracudas). Due to the high fish price in the local market, the fishers are driven to sell the big fish and consume the smaller ones.

There has been a declining catch yield of fish over the years. Fish species that occur in schools such as *Siganus spinus*, *Valamugil seheli*, *Rastrelliger kanagurta*, that used to be in abundance in catch yield 20 – 30 years back are very scarce nowadays (own observation, unpublished data). *Epinephelus polyphkadion* and *Epinephelus fuscogattatus* that aggregate annually to spawn is declining in numbers. The growing local demand for fish because of population increase, diet preference and political instabilities has led to the decline (Raj 2009). The artisanal catch landing of finfish for 2007 was approximately 4,418 MT and has decreased from approximately 6,240 MT in 2004.

Live Reef Fish Food Trade (LRFFT) that commenced in 1999 could also contributed to the decline of some of the targeted fish stock in the northern part (Bua province) of Fiji. The most targeted species includes Serranidae (coral trout, grouper and rock cod), and the Humphead

wrasse (*Cheilinus undulatus*) (Yeeting, 2001) with the estimated LRFFT stock in 2001 in the Bua area (northern Fiji) to be 3,750 tonnes.

Several studies have outlined the status of reef finfishes in inshore areas of Fiji. Jennings *et al.* (1995) suggested that Fijian reefs and their associated shallow water had the capacity to sustain total yields of reef associated fish of at least 3.4 tonne km<sup>-2</sup> qoliqoli year<sup>-1</sup> or 10.2 tonne km<sup>-2</sup> reef year<sup>-1</sup>. Kuster *et al.* (2005) estimated that over the 20 year period, the reef of Ono I Lau (East Fiji) have sustained a yield of between of 2.9 and 3.7 tonnes km<sup>-2</sup> year<sup>-1</sup> of reef-associated fish. Joined research between DOF and Society for the Conservation of Reef Fish Aggregation (SCRFA) revealed the scarcity of some of the large reef fish species such as the big Groupers (*Epinephelus polyphekadion*, *Epinephelus fuscogattatus*, *E. lanceolatus*), *Cheilinus undulatus* (Wrasse), and *Bolbometapon muricatum* (Parrotfish). Jennings *et al.* (2005) reported the declining in abundance of larger fishes in ten fishing grounds in Kadavu (south of Fiji). Dulvy and Polunin (2004a) also reported the status of *B. muricatum*, which is locally extinct in some islands. While Sadovy *et al.* (2006) highlighted the disappearance and close to extinction of some of the main reef fish spawning aggregation sites in Fiji.

The disappearance of the large reef fish species could possibly have an impact on the marine ecosystems. Fishing may lead to changes in the size, abundance and biomass (Jennings *et al.* 1995) thus; Jennings *et al.* (1996) suggested that the annual removal of 5% of fish biomass might cause structural changes in reef fish communities. The targeted species are mostly the top predators or the invertebrate feeders and piscivorous feeders. Studied conducted by Wilson *et al.* (2010) on coral reef fish communities in Fiji suggested that overfishing and habitat degradation through climate change posed as the biggest threats to sustainability of marine resources.

#### 1.4. On-going Data Collection

The Fiji Fisheries Department is the main government body that is responsible for the management, conservation and sustainability of marine resources in Fiji. Consequently, the department set up two projects to gather independent fisheries data and dependent fisheries data within the inshore area or *I Qoliqoli*. The Statistics Section within the Management Services Section is responsible for gathering dependent fisheries data by collecting catch data from the artisanal fishers. The data are gathered from major fish markets around the country; whilst, Marine Resource Inventory Survey Project (MRIS Project) within the Research Section is responsible on collecting independent fisheries data, by undertaking underwater visual census with divers recording fish and invertebrates sighted along 50 m transect lines.

The objective of the MRIS Project is to undertake fisheries resource inventory of all the *I Qoliqoli* in Fiji (i.e. a total of 410 delineated traditional marine area boundaries, which include the major rivers and streams). The project is to gather baseline biological, ecological and socio-economic information and data sets on fisheries resources that would allow the department to understand the health, productivity and biodiversity that exist within each traditional fishing boundary. It would help in the formulation of appropriate management plan for each *I Qoliqoli*. In addition to that, the information will also allow the department to promote:

- Better living standards and alternative livelihoods for local communities
- Maintenance of food security

- Appropriate conservation actions and strategies that need to be incorporated as a precautionary strategy, and using adaptive management approaches (to species, ecosystem-based and socio-economic level).
- Community based inshore resource conservation, planning and management and their social and technical empowerments (co-management arrangement, utilizing a bottom-up approach).

The MRIS Project employed two methods of collecting data that are socio-economic survey (prepared questionnaires for household interview) and biological survey (underwater visual census).

### 1.5. Purpose of the Study

This study is undertaken in the spirit of the FAO Code of Conduct for Responsible Fisheries that states that conservation and management decisions for fisheries should be based on the best scientific evidence available. This project provides an overview of the status of finfishes in 16 *I Qoliqoli* in Fiji. The study is to estimate the total yield of reef-associated fish in each *I Qoliqoli* surveyed by underwater visual census. The aims and objectives of this project were to:

1. Estimate population abundance, species assemblages' and composition.
2. Estimate the mean sizes and the size distribution within an *I Qoliqoli* (fishing grounds).
3. Estimate the existing stock of the most important food fish family in an *I Qoliqoli*.
4. Determine the Trophic Network of fish community in Fiji.

## 2. METHODOLOGY

Most research or sampling conducted in coral reef areas use underwater visual census (Brock 1982). Visual census is the best non-destructive method of population assessment in coral reef area, inexpensive and gives a rapid assessment of the area in limited time available. UVC provides a rapid estimation of relative abundance, biomass and length frequency distributions of reef fish. It involves the counting and estimation of fish sizes observed underwater by divers on a given transect (Yeeting 2001). It is based on onsite visual counts.

### 2.1 Underwater Visual Census

UVC is conducted along random paths (chosen by chance) along a 50 m x 5 m transect. A 50 m tape was placed on the substrate (sand, rubble or coral reef) to establish transect for each census carried out. Two divers were on the other side of the transect line, recording fish at 10 m interval (Figure 2). Therefore, the counting is done at every 10 m and a 5 m width from the transect line. Fish count, species identification and length (size) estimation were taken. Counting starts after 2 minutes in the water after the fishes are familiar with the presence of the divers. UVC method was also applied to surveying of invertebrates. Data were then recorded on a prepared data sheets.

Visual counts with scuba gears are used in deep part of the fishing ground (reef slope and channel) while snorkelling is conducted in shallow areas (lagoon and reef area).

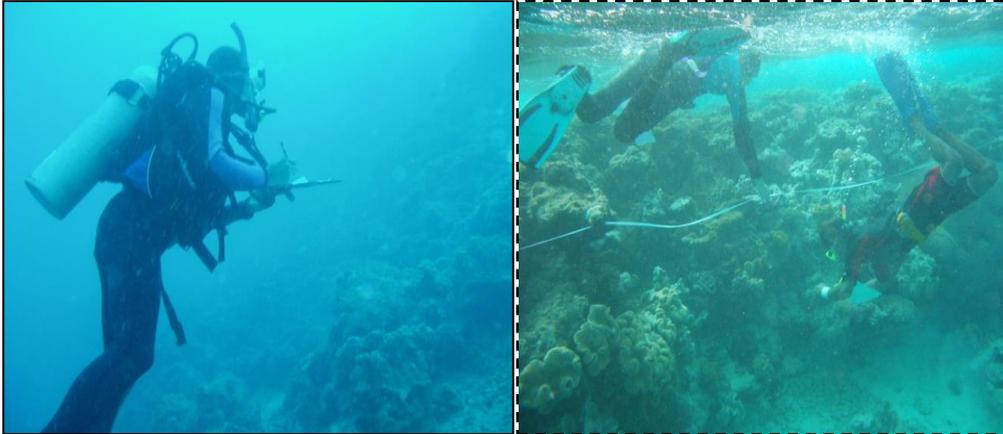


Figure 2: During visual counts in deep area/slope (left) and in shallow area (right) (DOF 2006).

## 2.2 Sampling Design

The objective of DOF is to survey 10–12 *I Qoliqoli* per year. The *I Qoliqoli* survey is randomly chosen by the department or sometimes upon request by the communities. The number of stations depends on the area of the *I Qoliqoli*, therefore each *I Qoliqoli* have different number of stations.

In this study, 16 *I Qoliqoli* from different areas of the Fiji group are analysed. The surveys were conducted from 2006–2008. Tabulated below are the survey areas with the number of stations (Table 1).

The number of transects are the same as the number of stations per fishing ground (1 transect/station). Each station represent different habitat within a fishing ground. All the fish that the divers come across are recorded (species name and estimated length).

Table 1: The *I Qoliqoli* surveyed, and number of transects. Geographical location within the Fiji Groups is shown in Figure 3. The areas are ordered according to the different fishing intensity (low to high intensity).

Areas	I Qoliqoli	Number of Transect
East Fiji [Lau Group]	Oneata Island	4
	Nayau	13
South Fiji [Kadavu]	Namuana	8
	Naikorokoro_Druwe	16
Central Fiji	Tikina Nasinu	10
	Tikina Levuka	5
North Fiji [Vanua Levu]	Karoko	3
	Vusasivo	4
Viti Levu		
Northern Viti Levu	Wananavu	5
	Rakiraki	4
Western Viti Levu	Tavua	30
	Ba	15
Southern Viti Levu	Galoa	12
	Namaqumaqua	8
Eastern Viti Levu	Mau	7
	Lami	5

## 2.3 Site Description

### 2.3.1. East Fiji (Lau)

Lau Islands are located on the far eastern part of the Fiji Group. Access to the island is by commercial or government vessels, which visit all islands once a month, depending on the availability of the vessel, or else by small airplanes once a week. The survey was conducted in Nayau Island and Oneata Island. The population of Nayau Island is approximately 500 people while Oneata is approximately 200 people.

Copra is the main source of income. Also the islanders are one of the best mat weavers and *masi* (traditional cloth use in traditional ceremony such as weddings, births etc.) makers selling their products in Suva (capital of Fiji). It is a lucrative business in Fiji. Few islands are involved in seaweed farming with the assistance of the Fiji Fisheries Department. The dried seaweeds are exported to Japan to be used as cosmetics and the pharmaceutical product. Still, subsistence farming and fishing is an important way of life in the island. If cash is needed then extra fish from the subsistence catch is sold to government officials who work in the islands.

Live Reef Fish Food Trade was conducted in Oneata Island, Lakeba Island and Bukatatanoa Reef in 2002 to 2003.

### 2.3.2. Central Fiji (Ovalau and Motoriki)

Lomaiviti Province is on central Fiji. The survey was conducted in the island of Ovalau, which lies off the eastern coast of Viti Levu, Fiji's main island. The population on Ovalau Island is approximately 3,000 people (Fiji Bureau of Statistics, 2009). The survey was undertaken in the *I Qoliqoli* of Tikina Levuka (11 villagers) and Tikina Nasinu (7 villagers). The fishing ground is located adjacently to each other.

Fiji's old capital, Levuka is on Ovalau. Whalers and missionaries founded Levuka in the 1830s. By the mid 19th century it was one of the main ports of call for trading ships and whalers throughout the South Pacific (UNESCO Report, 2000).

Tuna Cannery Company, Pacific Fishing Company (PAFCO) situated in Levuka provides employment to some of the islanders. Some in both islands practices small-scale artisanal fisheries and farming. The men planted *yaqona* (dried roots used for traditional drink in traditional ceremony but now as a social drink in every households), which is a lucrative business in Fiji. Fish caught is either sold in the village or taken to Suva and Levuka. However, subsistence fishing and farming is still an important way of life.

### 2.3.3. North Fiji (*Natewa and Tunuloa District*)

Vanua Levu (Fiji's second largest island) is on the northern part of Fiji, with a population of approximately 136,000 people (Fiji Bureau of Statistics, 2009). There are three provinces on this island. Great Sea Reef, third longest barrier reef in the world is located on the north coast (Figure 1). Some parts of the reef are being protected as a no take zone (no fishing and harvesting) by the local communities.

The survey was conducted in the *I Qoliqoli* of Vusasivo (2 villagers) and Karoko (1 village) with the population of approximately 600 and 300 people respectively, both in the province of Cakaudrove.

The main source of income is artisanal farming and fishing. Fish is mostly sold to middlemen who sell them in major fish markets in the North, Labasa and Savusavu Town. Gold Hold Seafood Company based in Labasa is the major buyer of seafood (fish and beachdemers) in the North. The beachdemers are exported while the fish are sold in Suva.

Live Reef Fish Food Trade (LRFFT) was also conducted in Bua province, but not in the *I Qoliqoli* surveyed. It occurred in Galoa and Tavea Island area between 1998 till 2004, but now only Tavea Island is involved (2008 until now).

### 2.3.4. South Fiji (*Kadavu*)

Kadavu is located in the southern part of Fiji with a population of approximately 10,000 people (Fiji Bureau of Statistics, 2009). The *I Qoliqoli* surveyed is located adjacently to each other on the leeward side of the island. The main sources of income are *yaqona* farming and artisanal fishing. Villagers rely on the sea when the *yaqona* plant is not ready to be uprooted and sold. *Yaqona* takes about 3–4 years to mature. Fish are either sold in Vunisea government station, middlemen in some of the villagers or else transported to Suva via the commercial vessel.

The island is just off the southern coast of Viti Levu and access to the island is by commercial vessel that goes once a week and by small planes that fly daily. Poachers from Suva, which may practise illegal fishing such as diving using scuba or some destructive methods, target the fishing ground heavily. Each village have fish wardens that police the fishing ground. However, it is not so successful since poachers come at night in high powered boats. The first marine reserve established and gazetted in Fiji is located in Waisomo *I Qoliqoli* on the Great Astrolabe Reef (Figure 1). Reef fish spawning survey in Kadavu, by DOF and SCARFA in 2008, identified five spawning aggregation sites for *Epinephelus polyphkadion*, *E. fuscogattatus* and *Plectropomus areolatus*.

### 2.3.5. Viti Levu

#### **North-western coast**

The *I Qoliqoli* surveyed are under the jurisdictions of the *Vanua Ba* and *Tilivabukuya* and *Tavua*. Both are located on the northwestern coast of Viti Levu (Figure 3). In the last census report (1996-2007) the overall population of Ba and Tavua are 18,526 and 2,388 respectively with the population of Indo Fijians decreasing significantly (Fiji Bureau of Statistics, 2009). The areas are dominated by sugarcane farms, which provide employment to the local during the harvesting season. Vatukoula Goldmine also employed the people of Tavua for over the years (1934 – 2006), which increased dynamite fishing in these areas. Mine workers smuggled out dynamites and sold to fishermen, which became a norm and a fast way to earn easy cash. It's not only killed more fish at one time and its habitat but also more and more people were seriously injured as a result of dynamite fishing. Consequently, the Chief of the vanua Tavua established an *I Qoliqoli* committee in 2003 to fight against dynamite fishing in the area. The Chief supported and funded the policing of dynamite fishing in his fishing ground. However, illegal fishing is still common in the *I Qoliqoli* of *Ba* and *Tilivabukuya*. Fishers used crowbars to break corals and rubbles to fetch for holothurians species and sell to Chinese buyers. There is still an occasionally use of dynamite to catch fish. Small-scale commercial fisheries dominate in this area. Fishermen often travel as far as the Yasawa Group, and Bua Province in Vanua Levu to have a good catch. There is no community based management policies that governs the fishing ground, maybe because of the clash of ownership, which results in poor governance of the fishing ground.

#### **South Coast of Viti Levu – Serua Province.**

In this area, the population has increased from approximately 15,000 in 1997 to approximately 18,000 in 2007 (Fiji Bureau of Statistics 2009). The main sources of income in this area are tourists, fishing and logging. Coastal development for tourist resorts is common. The Pacific Harbour Tourism Complex is located in the province. Also the forest area is common with the much-debated mahogany trees. Majority of the fishers are both subsistence and artisanal. The catches are mostly sold on the roadside or else taken to Sigatoka or Navua town. Aquarium fisheries are conducted between Culanuku and Wainiyabia fishing area and also recreational fisheries such as sport fishing, scuba, fish feeding and shark dive and feeding. They are all based in the Pacific Harbour Tourism Complex.

#### **South East Coast of Viti Levu**

The areas are near the capital city of Fiji, Suva. Suva, being the largest urban centre in the South Pacific, has an increase in industrial and urban growth. The total population of Suva and its peri-urban area (surrounding area) is approximately 74,000 and 11,000 respectively (Fiji Bureau of Statistics 2009). The waste effluent from the city and industry enters the sea indirectly/directly and are then carried by currents to the neighbouring fishing areas. The fishing grounds also accommodate people residing in Suva. Low-income earner in the town area uses the fishing ground for extra source of protein.

#### **North Coast of Viti Levu (Rakiraki District)**

The area is more laid back compared to other areas of Viti Levu. The artesian water company (Fiji Water) located within the vicinity of Rakiraki provides employment to some of the locals. A few small tourist resorts are found along this coastline. The population is around 30,000 but continued to decrease at a faster rate (Fiji Bureau of Statistics 2009). Majority of the people living in this area are subsistence fishers and small-scale commercial farmers.

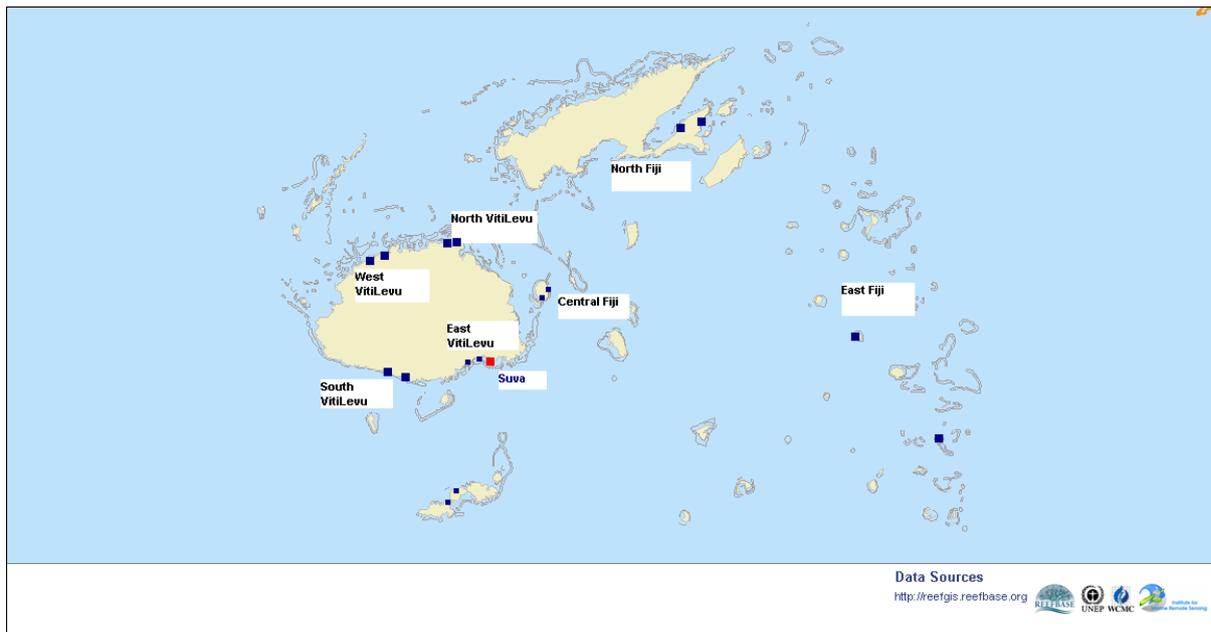


Figure 3: Location of the study sites (marked in blue) (Reef Base Online GIS).

## 2.4 Data Analysis

The data were analysed in Pasgear 2 software (version September 2008) (Kolding 2008), which is a customised database package primarily, intended for experimental or artisanal fishery, downloaded from [www.imr.no/forskning/bistandsarbeid/data/pasgear\\_2/en](http://www.imr.no/forskning/bistandsarbeid/data/pasgear_2/en). The software is useful for testing fishing data such as under water visual census (UVC) described by Labrosse *et al.* (2002). Before importing into the Pasgear, the species were given species identification codes. The species identification codes with coefficient a and b were provided by Dr. Jeppe Kolding of the University of Bergen. Species that were not in the list were given the identification code and the coefficient a and b were borrowed from the Fish Base website.

The software automatically estimates weights from length – weight relationships. Abundance is estimated from the observed number of species per transect (500m<sup>2</sup>).

$$D = \frac{\sum_{i=1}^p ni}{Ld}$$

where;

ni - number of fish seen

L – length of transect (50 m)

d – Width from the transect (5 m)

Biomass is calculated from the observed weight that is the number of fish multiplies by the weight from the length – weight relationship.

$$B = DW$$

where:

D - the density of fish per 500m<sup>2</sup> by the number of stations per fishing grounds

W – weight from the length – weight relationship.

Size distributions were determined on the length frequencies curve. The trophic network group is calculated in biomass and then imported to excel for the identification of species with the different trophic group.

The list of the important food fish species is obtained from the 'Food Fishes of Fiji' chart of the Fiji Fisheries Department. These are Groupers (*Plectropomus spp*, *Epinepheus spp*, *Cephalopholis spp*), Parrotfishes (*Chlorurus spp*, *Scarus spp*), Emperors (*Lethrinus spp*, *Gnathodentex spp*), Snappers (*Lutjanus spp*), Goatfishes (*Parupeneus spp*), Rabbitfishes (*Siganus spp*), Surgeonfishes (*Acanthurus spp*, *Ctenocahetus spp*), Barracudas, *Mugilidae spp*, *Hemiramphidae spp*, Tunas and Sweetlips (*Plectorhinchus spp*).

### 3 RESULTS

#### 3.1 Fish Abundance and Species Assemblages

A total of 544 fish species belonging to 55 families were recorded. The most diversified families are Pomacentridae (107 species), Labridae (63 species), Chaetodontidae (39 species), Acanthuridae (36 species) and Scaridae (35 species).

Damselfishes (Pomacentridae) and Surgeonfishes (Acanthuridae) had the highest fish counts, which were 9,827 and 4,204 respectively. Wrasses (Labridae) had high species diversity (66 species) but low fish count (1,402) as compared to Fusiliers (Caesionidae) that had low species diversity (12 species) and high fish counts (3,832). In addition, low numbers (1,412) of Butterflyfishes were observed but with high species diversity. The fish count and diversity (number of species) per family are shown in Table 7 in appendix 1.

##### 3.1.1 Density and Biomass of Finfishes per *I Qoliqoli*

Abundance and biomass of finfishes per 500 m<sup>2</sup> in each fishing grounds (*I Qoliqoli*) varies (Table 2 and 3). Of all the fishing grounds surveyed, Lami in East Viti Levu had the highest fish abundance (504/500 m<sup>2</sup>) and biomass (135.1 kg/500 m<sup>2</sup>) followed by Nayau (Lau Group) in East Fiji. Of all areas, Namaqumaqua (South Viti Levu) had the lowest fish abundance (43.3/500 m<sup>2</sup>) and fish biomass (7.0 kg/500 m<sup>2</sup>).

Table 2: Mean abundance and biomass estimates for each *I Qoliqoli* (per 500 m<sup>2</sup>), in outer islands fishing grounds (*I Qoliqoli*).

	East Fiji		South Fiji		Central Fiji		North Fiji	
	Nayau	Oneata	Naikorokoro	Namuana	Tikina Levuka	Tikina Nasinu	Vusasivo	Karoko
Abundance (No/transect)	466.7± 232.8	110.8± 165.1	237.4± 131.3	288.5± 101.9	100± 26.9	80.2± 15.5	76.0± 15.6	255.5± 187.4
Biomass (W(kg)/transect)	124.4± 74.9	11.9± 13.9	60.4± 68	52.9± 28.9	71.4± 70.2	18.4± 7.9	2.9± 3.1	19.9± 5.3

Table 3: Mean abundance and biomass estimates for each *I Qoliqoli* (per 500 m<sup>2</sup>) in main island, Viti Levu.

	North Viti Levu		West Viti Levu		South Viti Levu		East Viti Levu	
	Rakiraki	Wananavu	Ba	Tavua	Namaqu maqua	Galoa	Mau	Lami
Abundance (No/transect)	315± 169.7	199.5± 3.5	299.7± 117.1	187.2± 202.6	43.3± 18.9	159± 122.5	186.4± 131	504± 215
Biomass (w(kg)/transect)	64.6± 33	42.3± 4.3	21.3±1 0.3	43.2± 54.4	7.0± 3.5	17.3± 17.8	26.5± 25	135.1± 125.8

### 3.1.2 Abundance and Biomass per Habitat

The average abundance was highest in the lagoons but the variance was also high. The abundance of fish in lagoon, reef area, reef slope, back reef and fore reef were somewhat similar but different from the reef channel and outer reef (Figure 4). The biomass varied but was highest in the fore reef and lagoon but lowest in back reef and reef channel (Figure 5). East Fiji (Lau Group) (Figure 6 & 12): In lagoon and back reef area, more fish with high biomass were sighted in Nayau Island than Oneata. The fore reef of Oneata Island was not surveyed because of bad weather (strong currents).

South Fiji (Kadavu Island) (Figure 7 & 13): High fish abundance was seen in Namuana reef area but low biomass. While, high fish biomass was observed in Naikorokoro reef slope with low abundance.

Central Fiji (Ovalau Island) (Figure 8 & 14): Tikina Levuka had high abundance and biomass in all areas.

North Fiji (Vanua Levu) (Figure 9 & 15): Karoko had high abundance and biomass.

Viti Levu: Rakiraki had high abundance and biomass than Wananavu, while Ba had high abundance with low biomass and Tavua had low abundance with high biomass (Figure 10 & 16). In south Viti Levu (Figure 11 & 17), Galoa had high abundance and high biomass compared to Namaqu maqua. Lami had high abundance and biomass than Mau.

The abundance and biomass among *I Qoliqolis* varies. The number of transects observed were not equal due to the different area demarcated for each village/community. However, in areas less than 5 m depth the mean sizes of fish were less than 15 cm (Figure 26), which were mostly juvenile fish. Location of the fishing ground influences the distribution of fish within the habitat (Figure 18 – 24). Fishing grounds in outer islands had high mean abundance and

biomass in the lagoon than the main islands. The abundance and biomass of fish per habitat among *I Qoliqoli* show less variation.

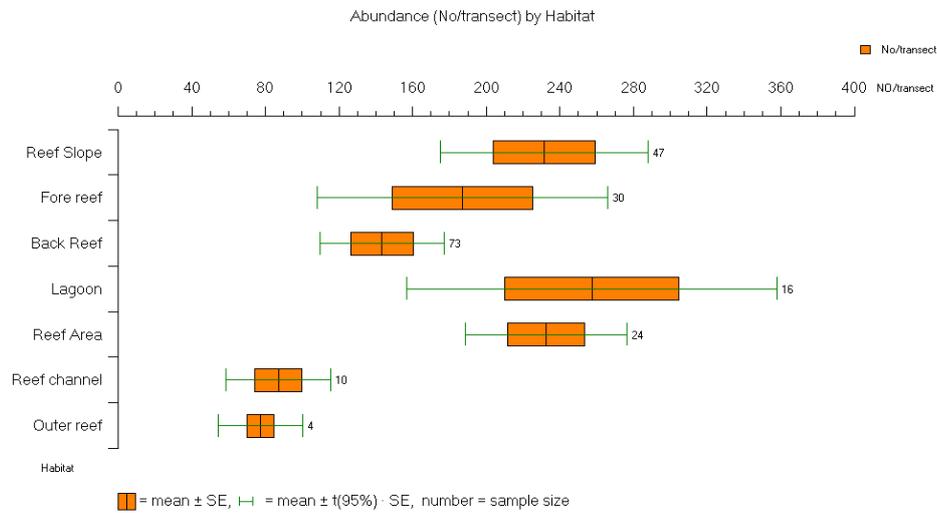


Figure 4: Fish abundance by habitat in all *I Qoliqoli*.

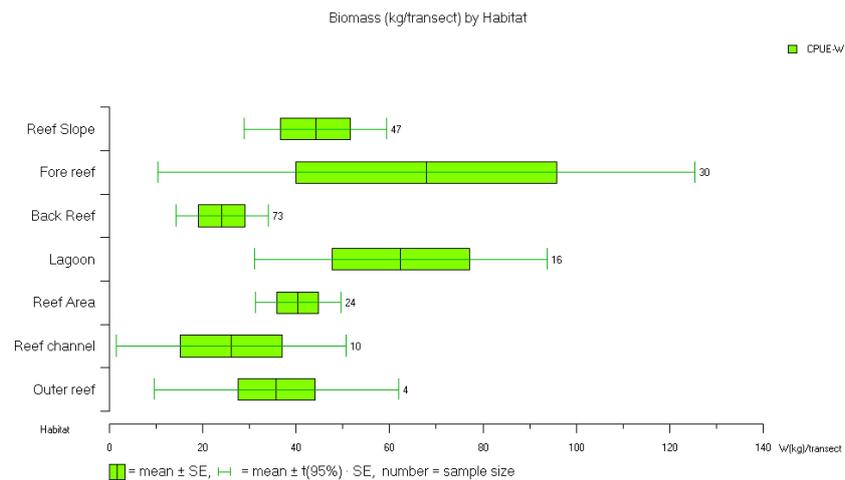


Figure 5: Fish biomass by habitats in all *I Qoliqoli*.

Overall, in the fore reef, Lami (East Viti Levu) has the highest fish abundance of mean (629.5/ 500 m<sup>2</sup>) and with the mean biomass of 61.9 kg/500 m<sup>2</sup>, followed by 463.5/ 500 m<sup>2</sup> in Nayau Island (East Fiji) and with a mean biomass of 323.1 kg/500 m<sup>2</sup>.

Lami and Nayau Island have also the highest mean fish abundance and biomass in the back reef area. High mean fish abundance was also sighted in the lagoon and reef channel area of Nayau Island. The reef area and reef slope (drop off) of Namuana has high mean fish abundance.

### Fish Abundance by Habitat in *I Qoliqoli*

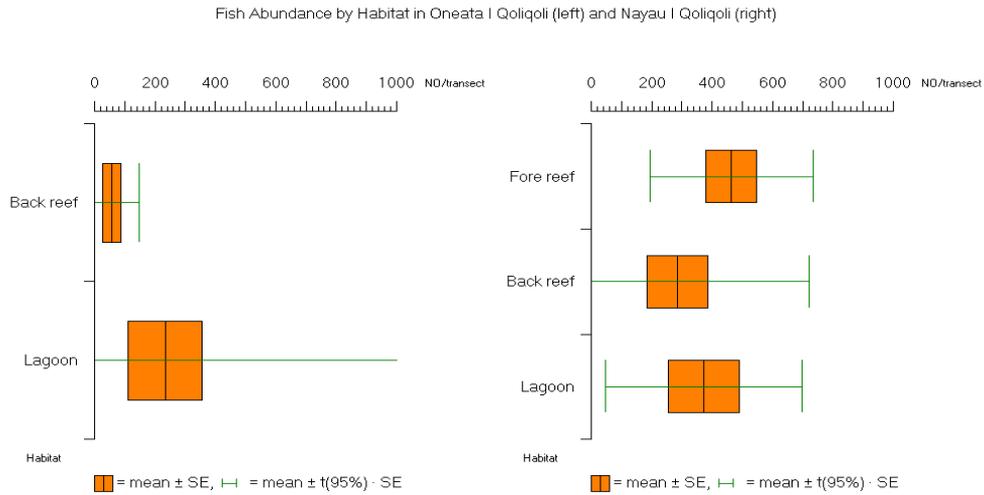


Figure 6: Fish Abundance (No/500m<sup>2</sup>) by habitat in Nayau Island and Oneata Island, East Fiji.

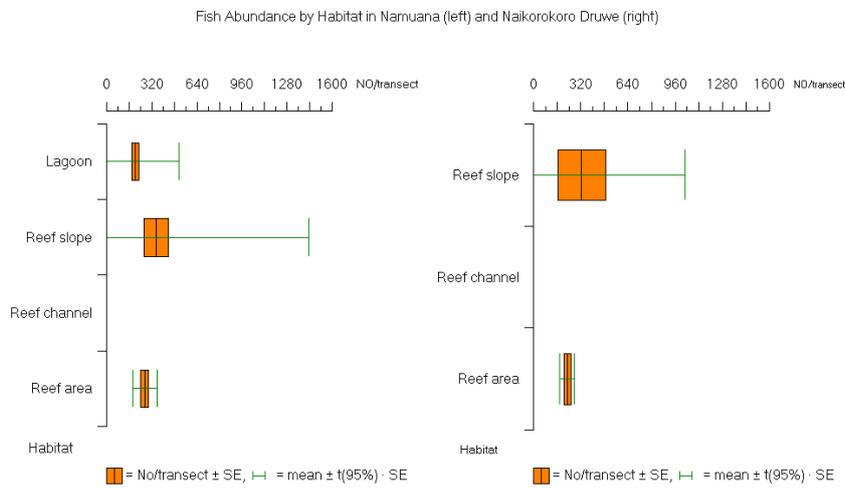


Figure 7: Fish Abundance (No/500m<sup>2</sup>) by habitat in Namuana and Naikorokoro, Kadavu Island, South Fiji.

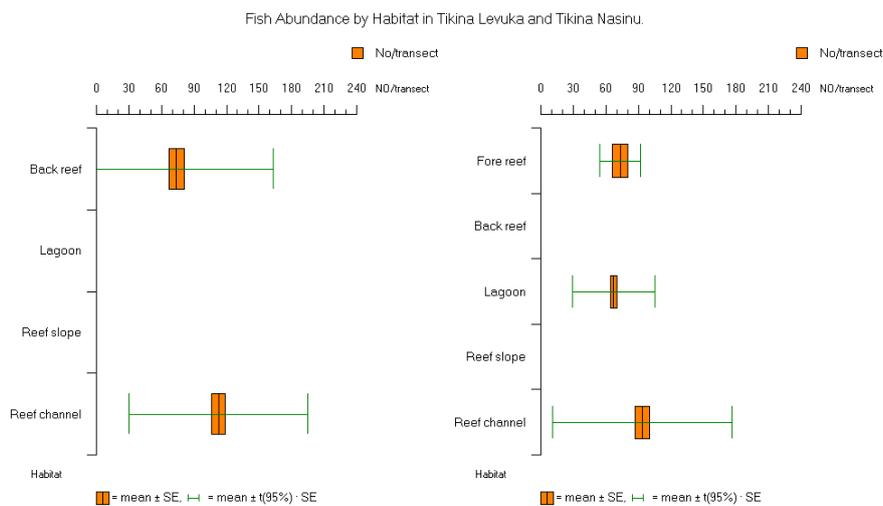


Figure 8: Fish Abundance (No/500m<sup>2</sup>) by habitat in Tikina Levuka (left) and Tikina Nasinu (right), Ovalau Island, Central Fiji.

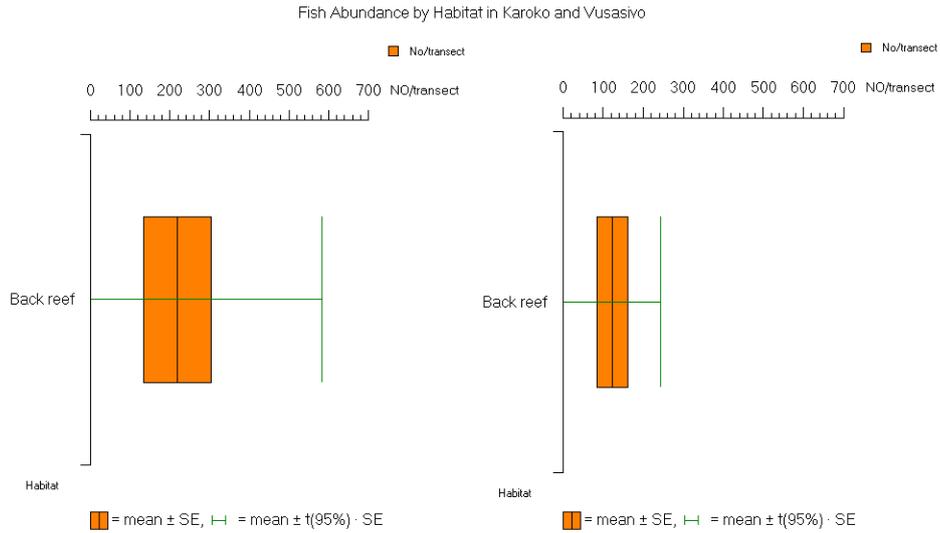


Figure 9: Fish Abundance (No/500m<sup>2</sup>) by habitat in Karoko (left) and Vusasivo (right), Vanua Levu, North Fiji.

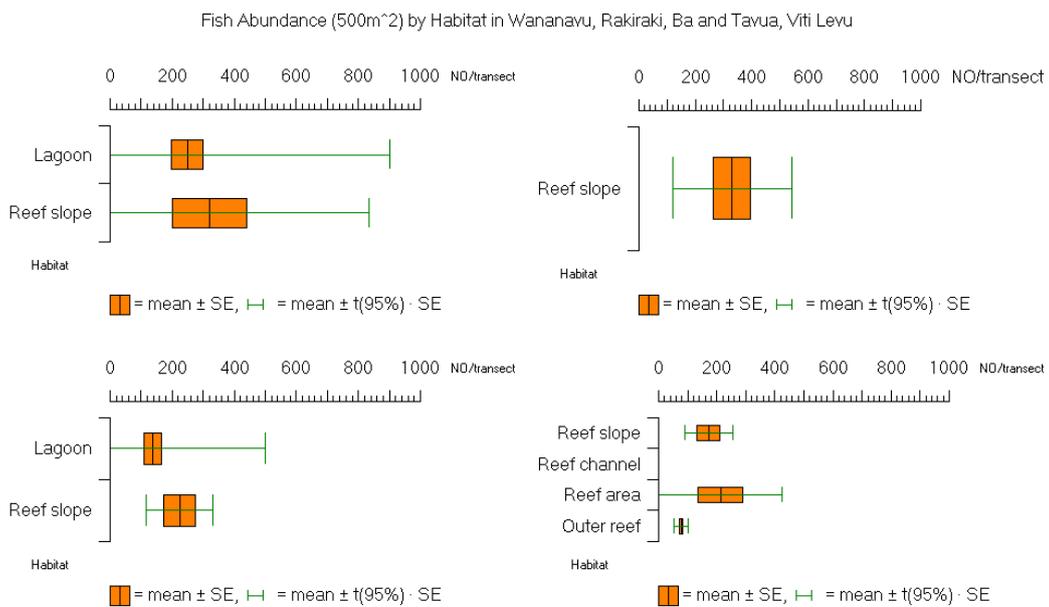


Figure 10: Fish Abundance (No/500m<sup>2</sup>) by habitat in Wananavu (top left), Rakiraki (top right), Ba (bottom left) and Tavua (bottom right), Viti Levu.

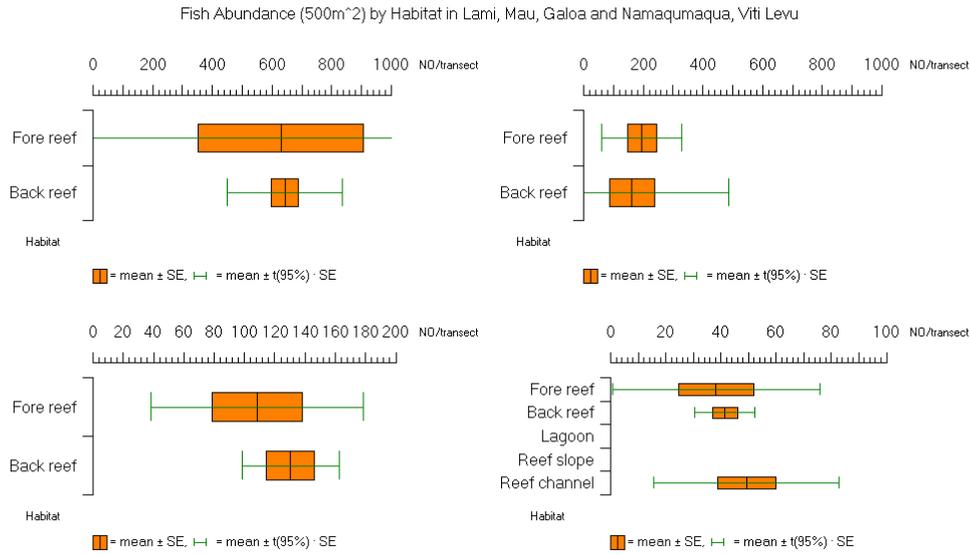


Figure 11: Fish Abundance (No/500m<sup>2</sup>) by habitat in Lami (top left), Mau (top right), Galoa (bottom left) and Namaqumaqua (bottom right), Viti Levu.

**Biomass by Habitat in *I Qoliqoli***

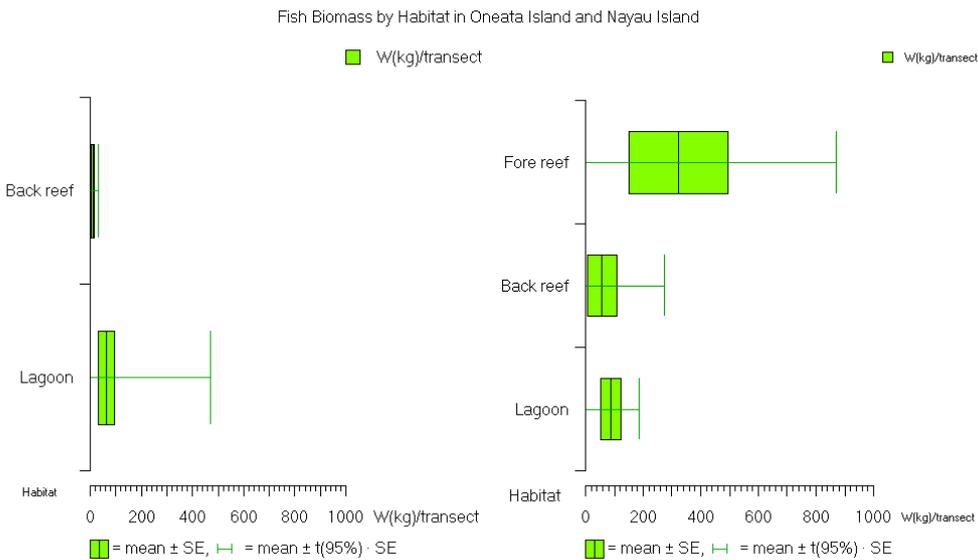


Figure 12: Fish Biomass (W (kg)/500m<sup>2</sup>) in Oneata Island (left) and Nayau Island (right), Lau group, East Fiji.

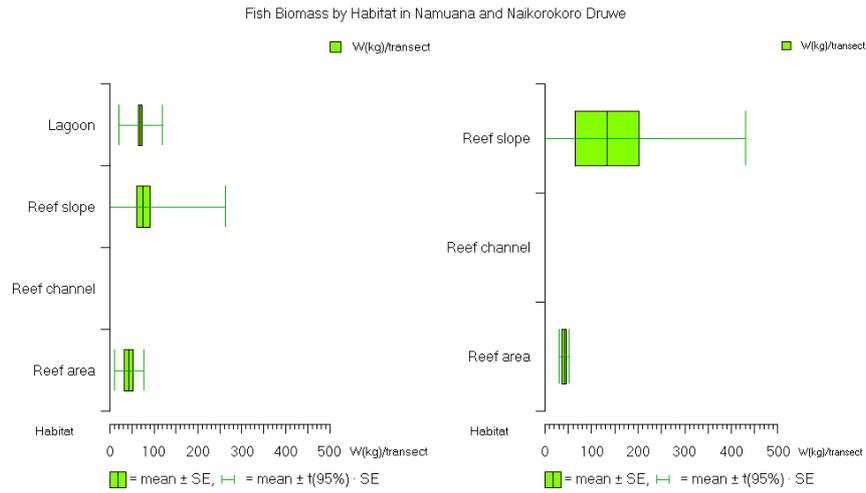


Figure 13: Fish Biomass ( $W(kg)/500m^2$ ) in Namuana (left) and Naikorokoro Druwe (right).

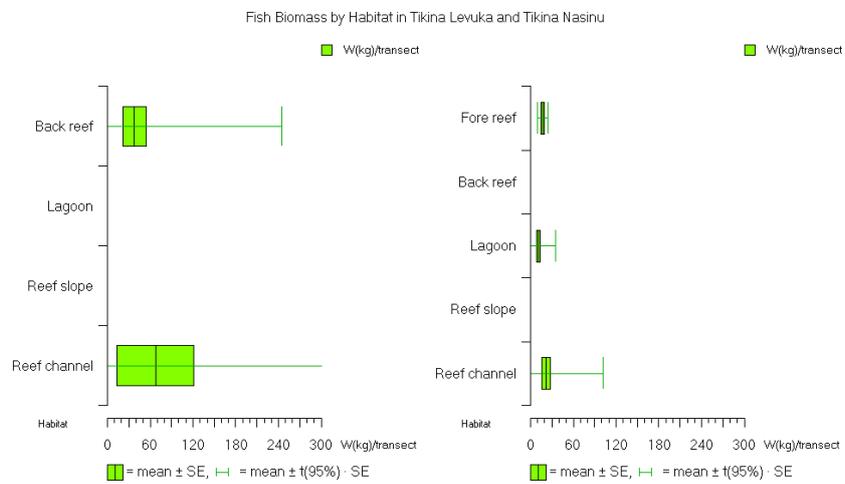


Figure 14: Fish biomass ( $W(kg)/500m^2$ ) in Tikina Levuka (left) and Tikina Nasinu (right), Ovalau Island, Central Fiji.

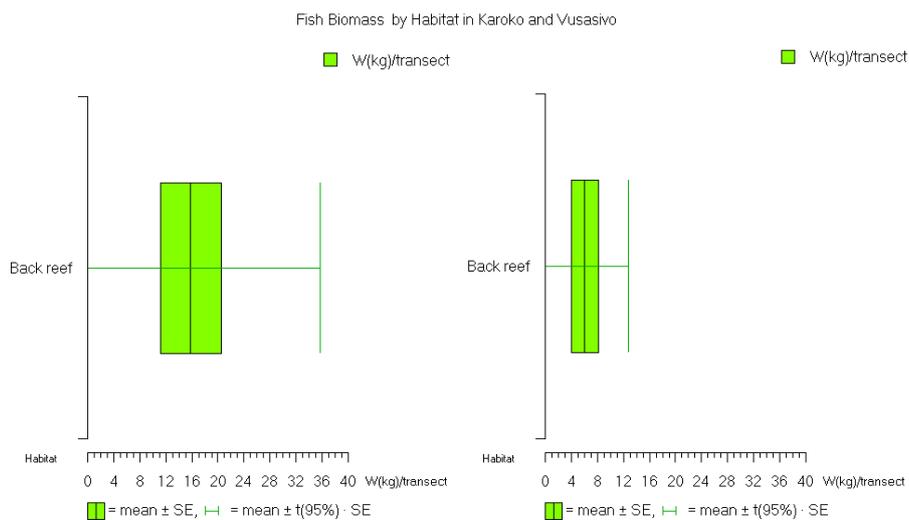


Figure 15: Fish biomass ( $W(kg)/500m^2$ ) by habitat in Karoko (left) and Vusasivo (right) in Cakaudrove, Vanua Levu, North Fiji.

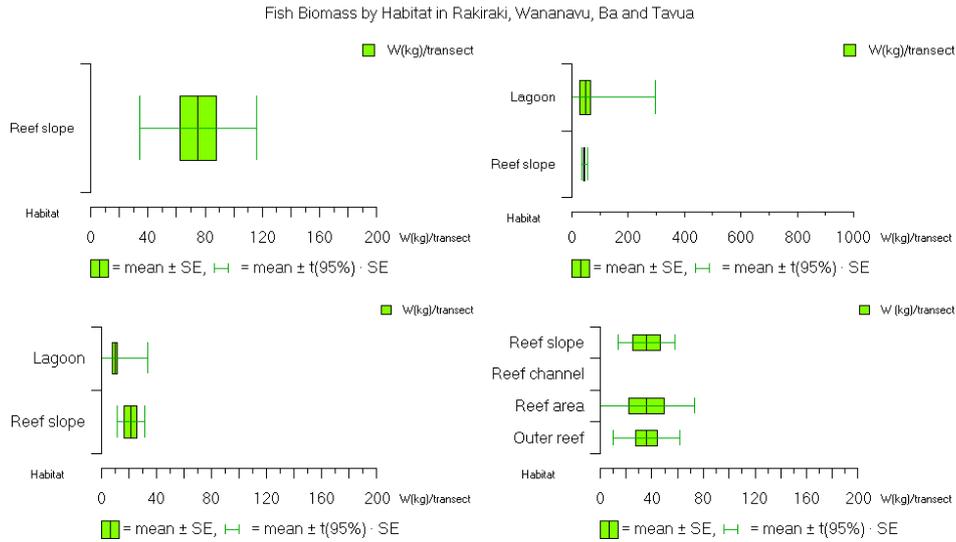


Figure 16: Fish biomass (W (kg)/500m<sup>2</sup>) by habitat in Rakiraki (top left), Wananaavu (top right), Ba (bottom left) and Tavua (bottom right), Viti Levu.

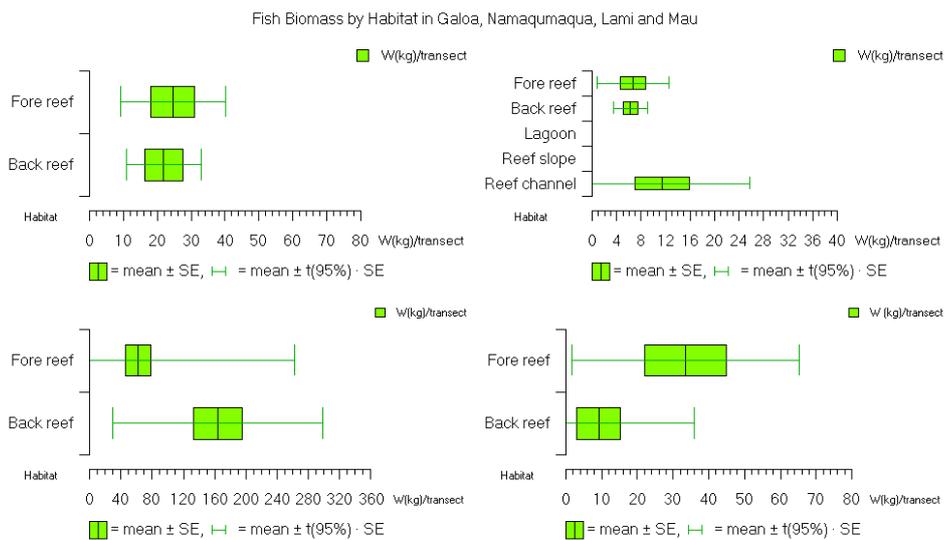


Figure 17: Fish biomass (W (kg)/500m<sup>2</sup>) in Galoa (top left), Namaqumaqua (top right), Lami (bottom left) and Mau (bottom right), Viti Levu.

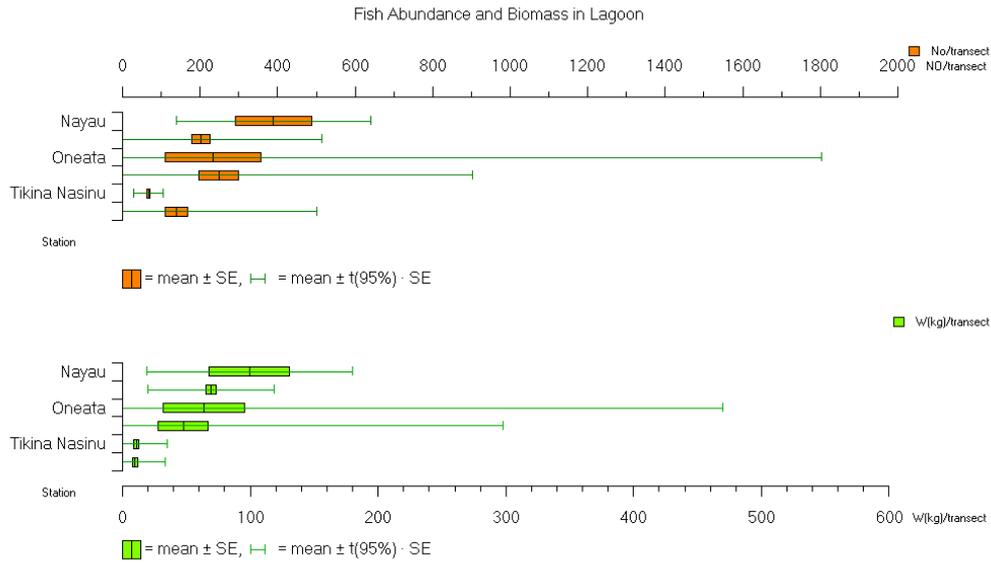


Figure 18: Fish abundance (No/transect) and biomass (kg/transect) in lagoon area.

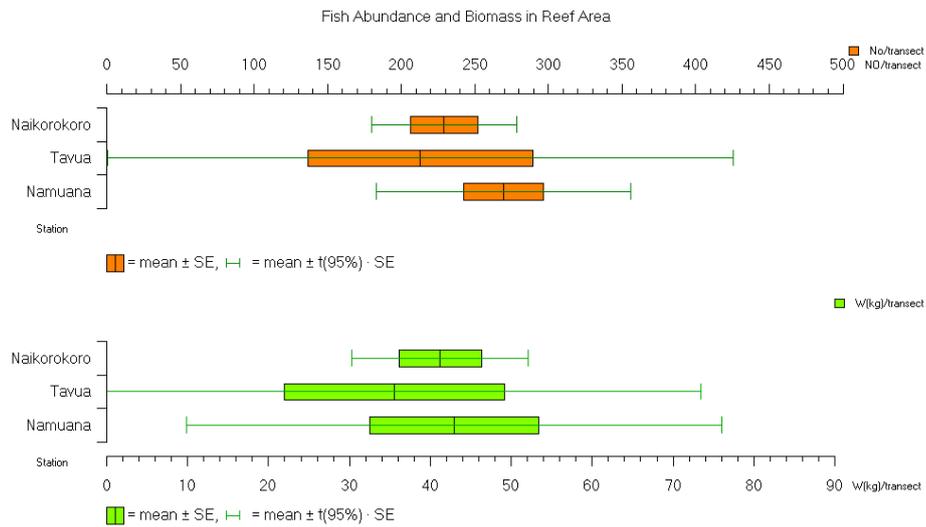


Figure 19: Fish abundance (No/transect) and biomass (kg/transect) in reef area.

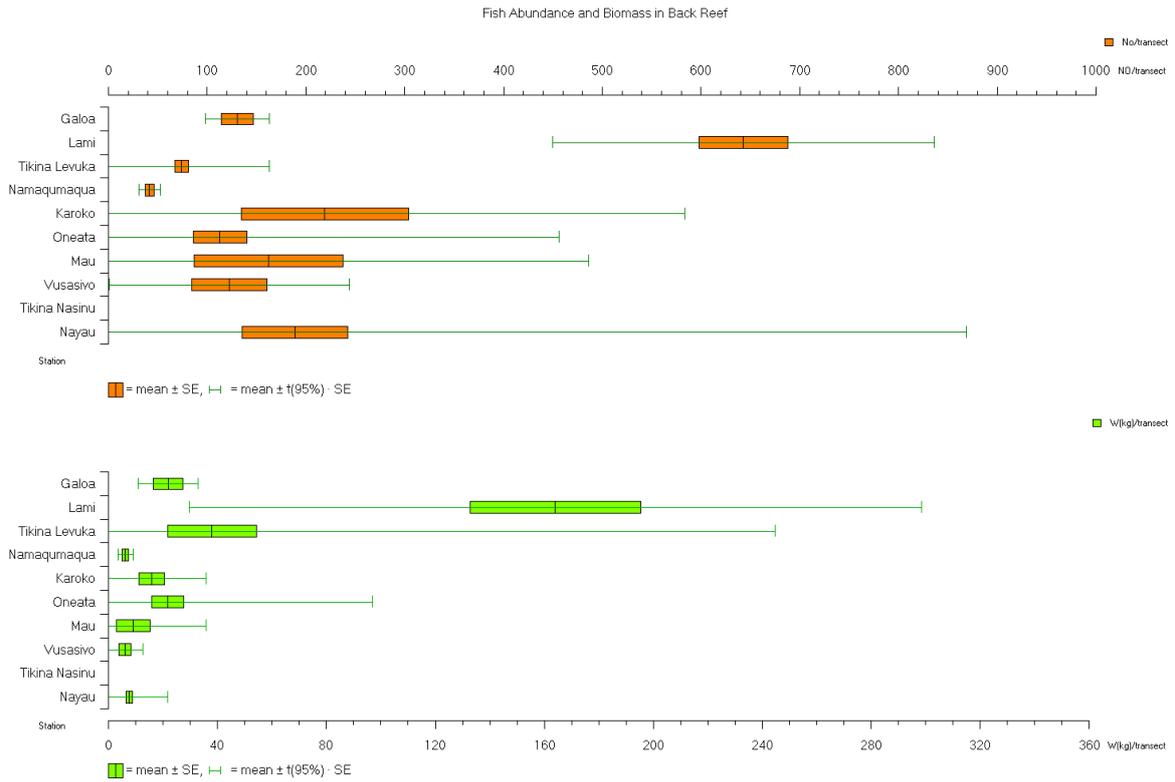


Figure 20: Fish abundance (No/transect) and biomass (kg/transect) in back reef.

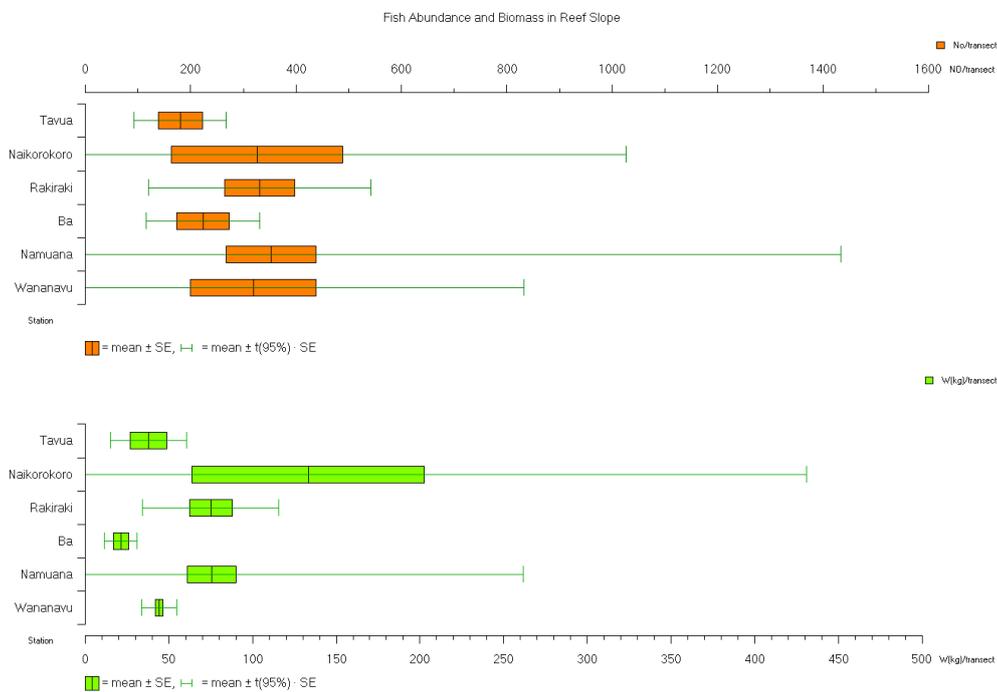


Figure 21: Fish abundance (No/transect) and biomass (kg/transect) in reef slope.

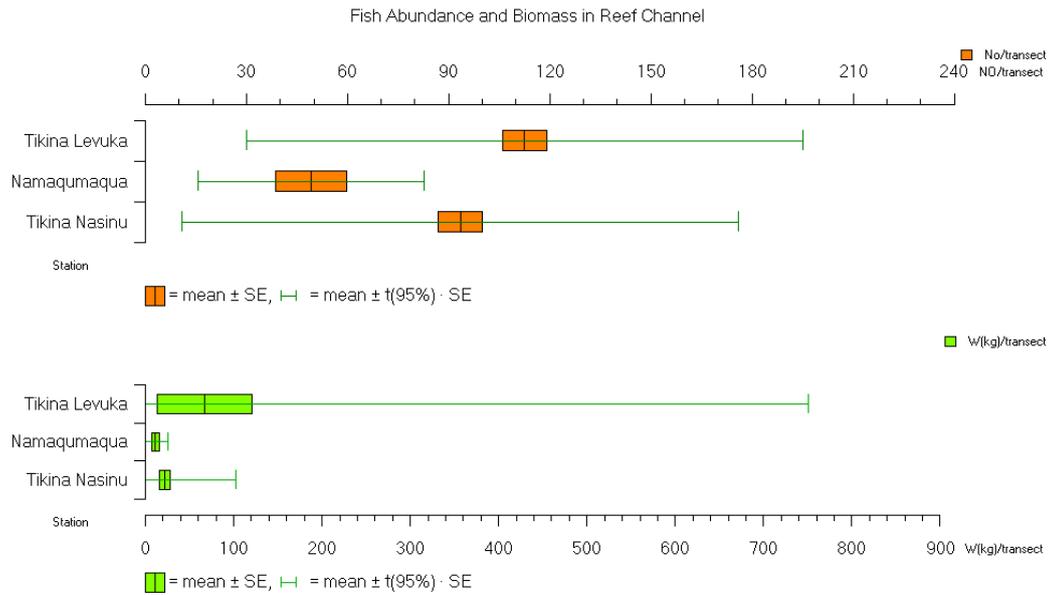


Figure 22: Fish abundance (No/transect) and biomass (kg/transect) in reef channel.

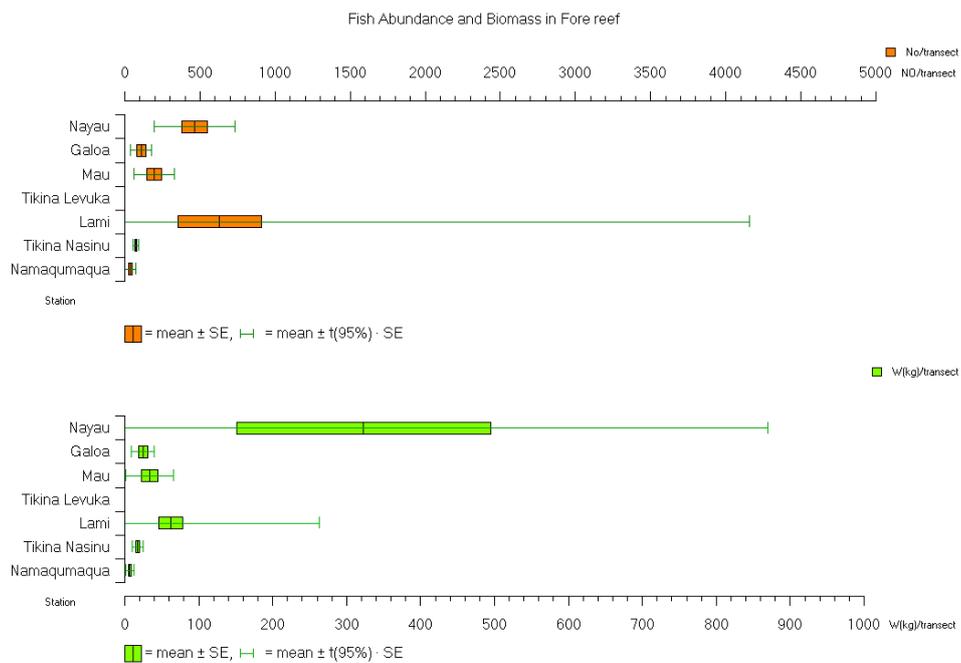


Figure 23: Fish abundance (No/transect) and biomass (kg/transect) in fore reef.

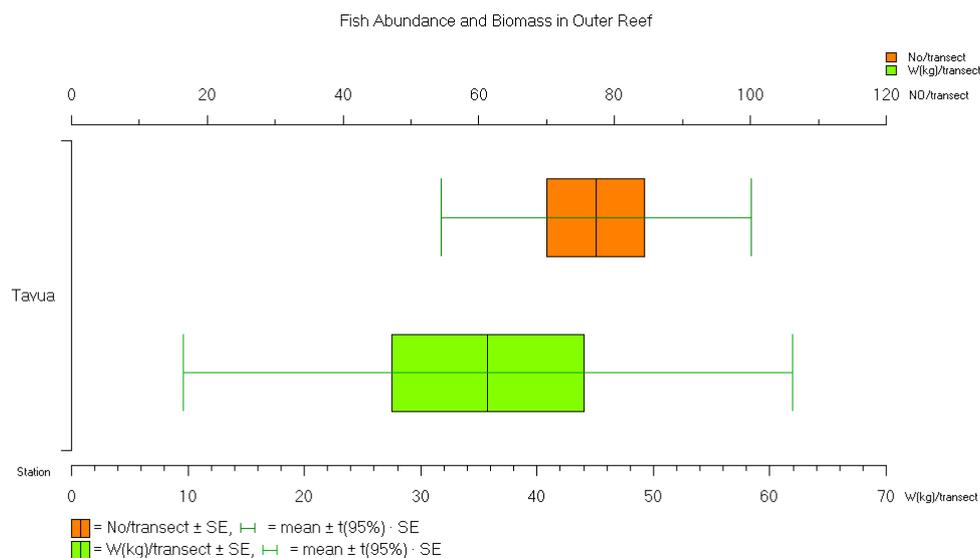


Figure 24: Fish abundance (No/transect) and biomass (kg/transect) in outer reef.

### 3.1.3 Abundance and Biomass of 5 Important Food Fish by Habitat

The 5 most important food fish families (commonly used as both subsistence and commercial) are shown in table 4, with Lutjanidae (Snapper), Lethrinidae (Emperor) and Serranidae (Grouper) listed as highly commercial. All were observed in all habitats except for Serranidae (Groupers), which was not sighted, in the outer reef (Table 4), probably because of the behaviour of the fish, preferring to hide and feed among corals. Groupers are rarely found in schools and only aggregate when it is time to spawn on reef channels and promontories. High mean abundance of groupers was sighted in lagoon. These are *Epinephelus merra* which commonly seen in shallow coral reef area. Also juvenile groupers are found closer to the shore to seek protection until they reach maturity.

High mean abundance and biomass density of Lutjanidae (Snappers), Lethrinidae (Emperor), Mullidae (Goatfish) and Scaridae (Parrotfish) were seen in lagoon area, while high mean abundance and density biomass of Acanthuridae (Surgeonfish) were found in the reef area (Table 4 and 5).

Tikina Nasinu (Central Fiji) had high mean abundance (9.4/transect) and biomass density (2 kg/transect) of Lethrinidae, while high mean abundance (87.3/transect) and biomass (14.4 kg/transect) of Scaridae were observed in Namuana. The other food fish have generally the highest mean abundance and biomass in Nayau Island (Table 8 and 9 in appendix 2a and 2b).

Table 4: Abundance (No/transect) of 5 important food fish by habitat.

Family	Lagoon	Back Reef	Reef Area	Reef Slope	Reef Channel	Fore reef	Outer reef
Lutjanidae	23.9±44	8.2±25.7	8±16	22.1±50.3	11.9±21.4	11.2±20.3	4.3±3.2
Lethrinidae	12.5±19.8	2.6±5.2	5±4	2.4±9.9	4.8±7.7	6.8±9.9	0.3±0.5
Serranidae	1.9±2.3	1.3±5.8	0.7±1.1	0.7±1.0	0.4±0.5	1.5±2.2	NULL
Scaridae	33.5±26.1	8.9±10.9	23.5±16.4	30.1±43	8.1±7.5	21.2±30.1	17.8±5.1
Acanthuridae	44.9±42	21.6±48.5	52.4±37.8	12.8±23.8	15.9±10.1	30.5±45.6	13.8±4.9
Mullidae	11.6±27.6	4.3±8.3	10.8±10.9	2.9±6	8.3±12.4	8.6±13.7	1.3±1.5

Table 5: Biomass (kg/transect) of 5 important food fish by habitat.

Family	Lagoon	Back Reef	Reef Area	Reef Slope	Reef Channel	Fore reef	Outer reef
Lutjanidae	13.3±24.6	2±6.8	1.5±2.6	4.1±8.2	9.6±24.4	4.5±8.9	1.1±1.4
Lethrinidae	6.1±15.4	0.7±1.6	0.9±1	0.8±2.7	2.2±2.8	4.2±11.9	0.2±0.3
Serranidae	0.8±1.1	0.2±0.5	0.2±0.8	0.3±0.8	0.2±0.2	0.8±1.7	NULL
Scaridae	11.4±9.5	3.1±5.3	7.5±8.8	7.2±6.7	2.5±2.6	9.9±15.6	16.5±10.9
Acanthuridae	10.6±11.1	5.8±15.4	10.8±7.5	3.7±8.7	3.7±3.2	7.7±12.5	6.7±2.5
Mullidae	7.3±23.4	0.8±1.7	3.1±7.7	0.7±1.6	2±2.9	2.2±3.5	0.5±0.8

### 3.2 Size Distribution by *I Qoliqoli*/Area Habitat and Depth

The mean length in outer reef is higher than the other habitats but it also has the lowest number of observation. There is no big difference in the mean lengths observed in other habitats (Figure 25).

In general, bigger fishes are found in the deeper part of the fishing grounds. Between 10 m and 25 m the dominant mean sizes are from 20–25 cm. However, some fish of these sizes (20 – 25 cm) are found in the shallow part (4 m – 10 m) of the fishing grounds (Figure 26).

Figure 27 – 34 illustrates the size distribution in an area. Steepen slope probably due to high fishing pressure (with the large fish removed), or more small fish in an *I Qoliqoli* (smaller community).

Less than 10 cm fish were fully observed in East Fiji (Figure 27), North Fiji (Figure 30), South Viti Levu (Figure 32) East Viti Levu (Figure (Figure 34) and West Viti Levu (Figure 33), thus this may be due to the high abundance of juvenile fish present. The fully observed fish in Central Fiji (Figure 28) and South Fiji (Figure 29) were bigger than 15 cm length, which probably due to its medium to large community. The increased in biomass and abundance in the size classes (11 – 25 cm), suggested some form of compensatory indirect effect of exploitation (Dulvy *et al.*, 2004a).

The steeper slope shown by West Viti Levu (Figure 33) may represent the increase in fishing pressure in the area. According to Dulvy *et al.*, (2004a) steepend slope of community size spectra represents exploitation.

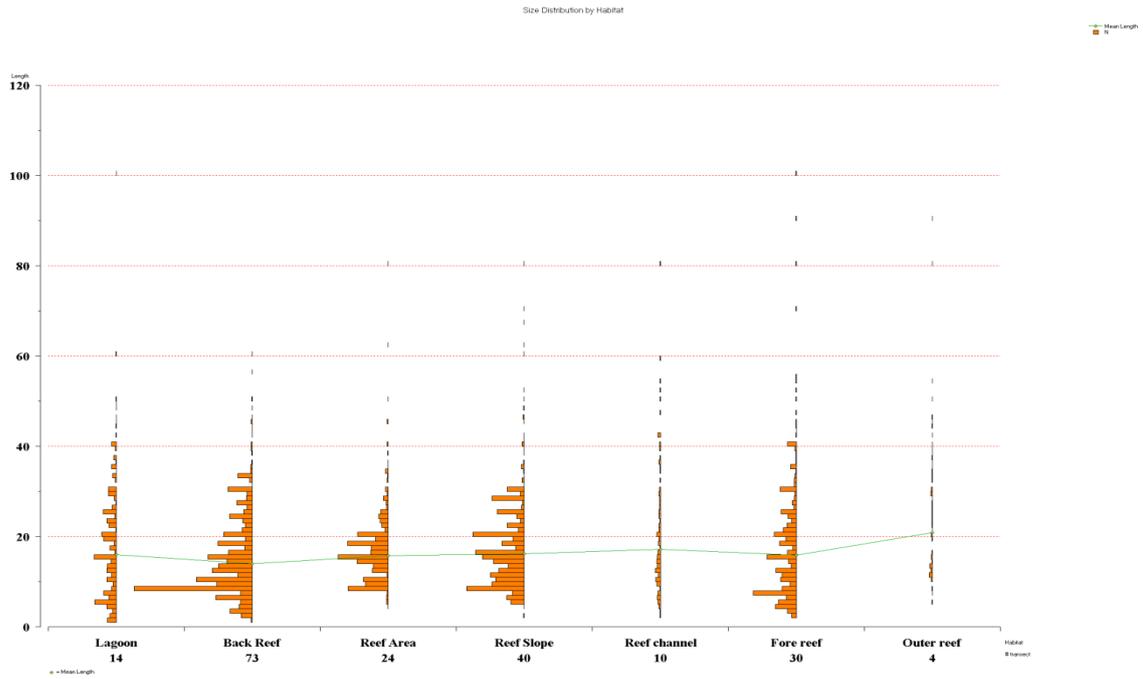


Figure 25: Size distribution of fish by habitat.

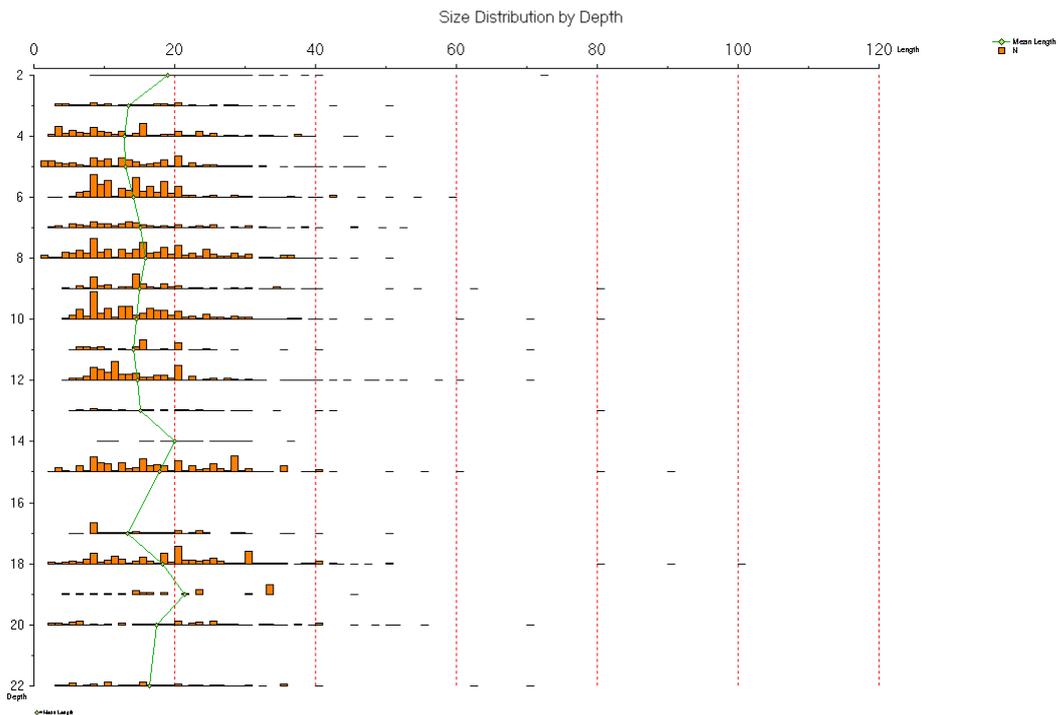


Figure 26: Length distribution by depth.

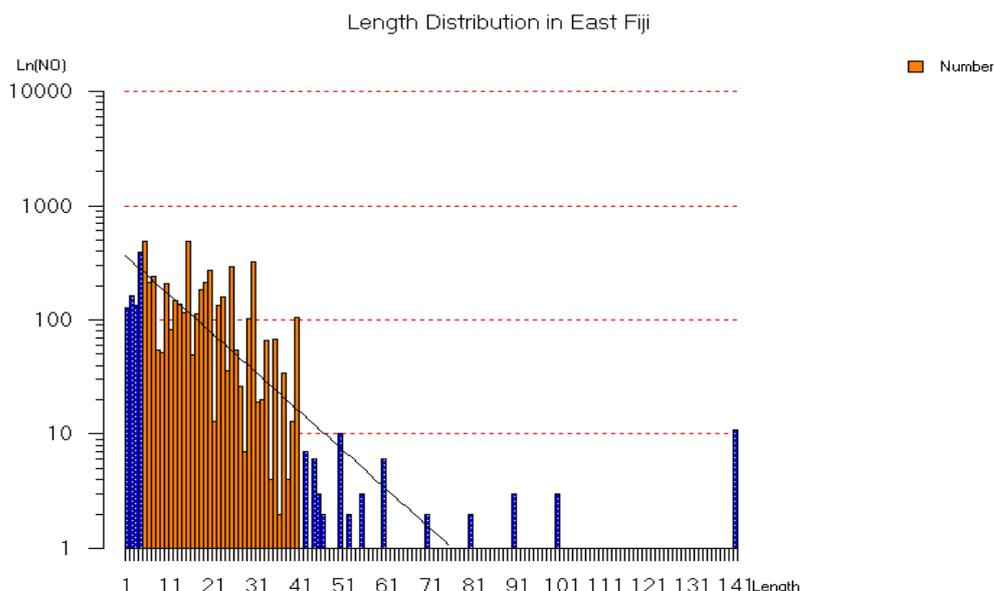


Figure 27: Length distribution in Oneata and Nayau Island, East Fiji. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

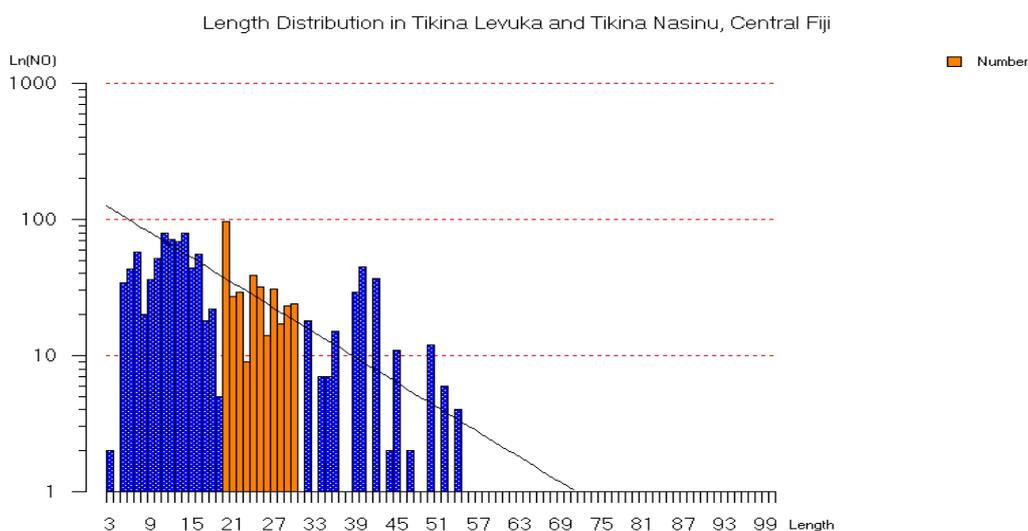


Figure 28: Length distribution in Tikina Levuka and Tikina Nasinu. The *I Qoliqoli* is located adjacently to each other. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

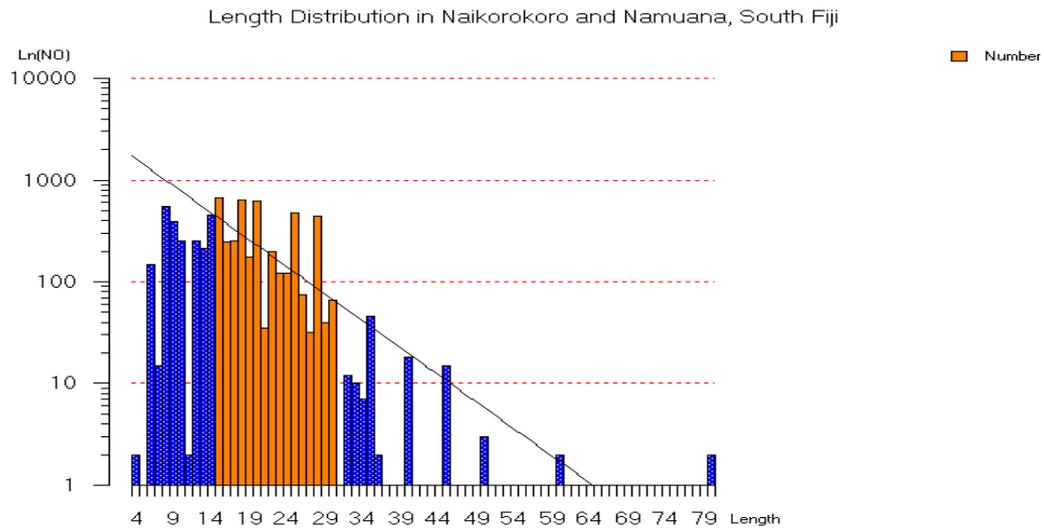


Figure 29: Length distribution in Naikorokoro and Namuana, South Fiji. The *I Qoliqoli* is located adjacently to each other. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

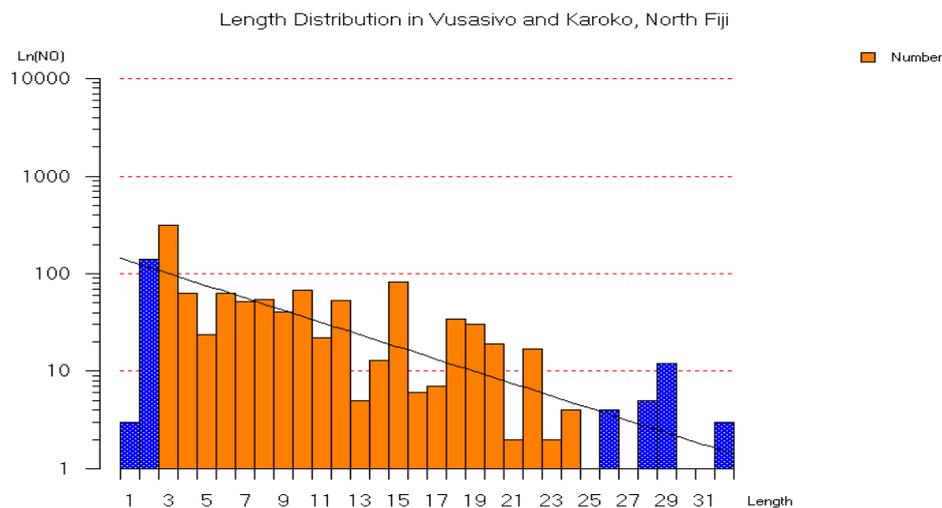


Figure 30: Length distribution in Vusasivo and Karoko, North Fiji. The *I Qoliqoli* is located along the same coastline in Natewa district. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

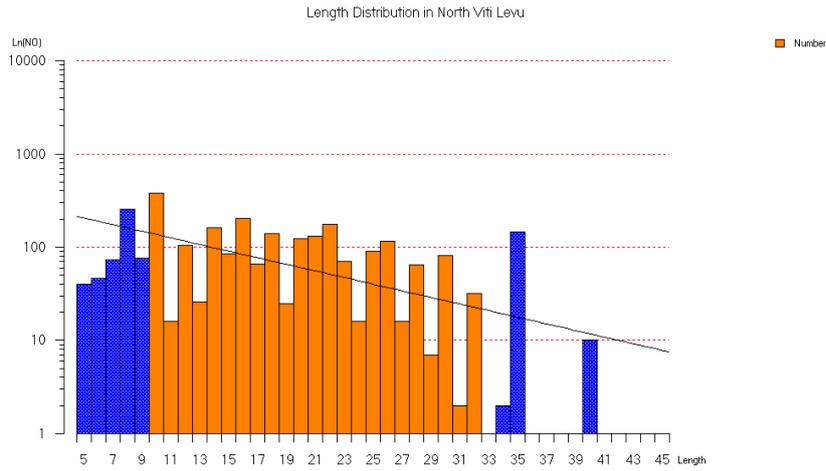


Figure 31: Length distribution in Wananavu and Rakiraki, North Viti Levu. The *I Qoliqoli* are located adjacently. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

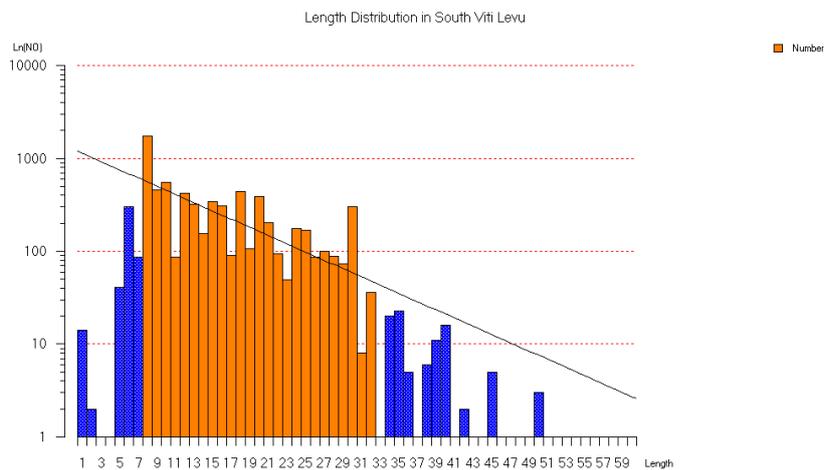


Figure 32: Length distribution in Galoa and Namaqumaqua, South Viti Levu. The *I Qoliqoli* is located on the same coastline. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

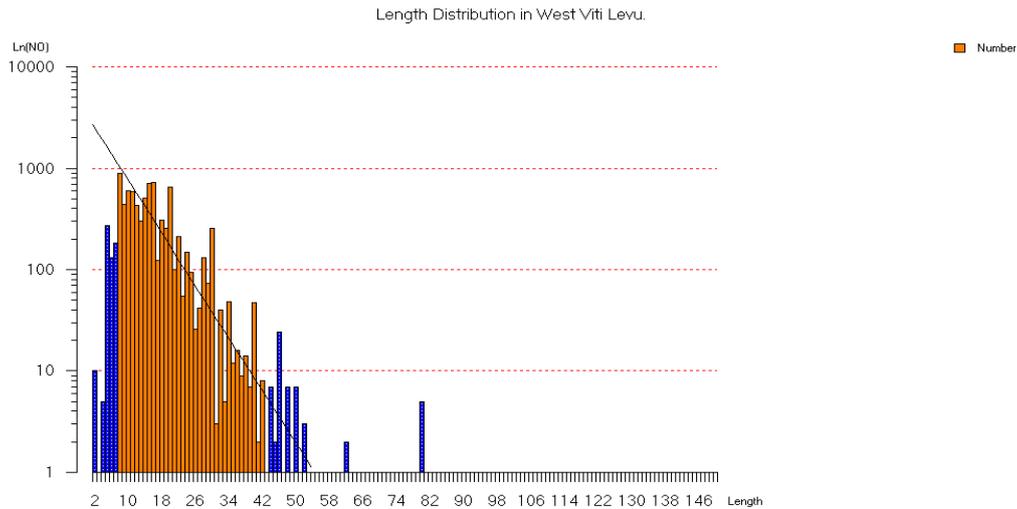


Figure 33: Length distribution in Ba and Tavua, West Viti Levu. The *I Qoliqoli* is located adjacently. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

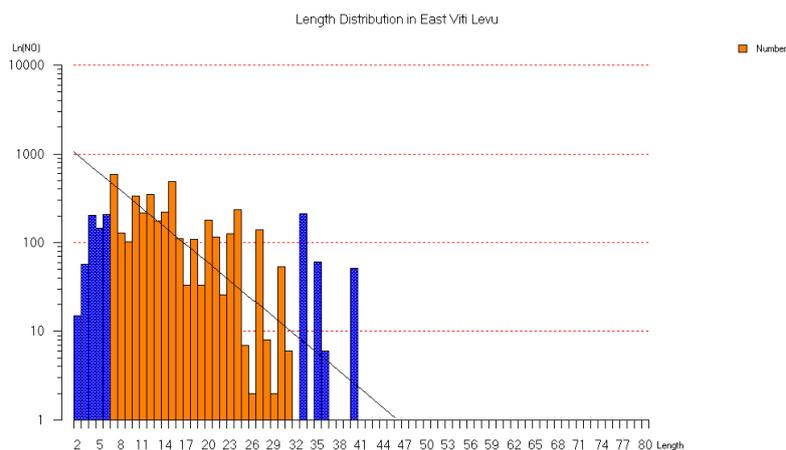


Figure 34: Length distribution in Lami and Mau, East Viti Levu. The *I Qoliqoli* is located on the same coastline. The blue bars are not fully represented (the idea is that probably some fishes were not observed). Orange bar is fully represented or observed.

### 3.3 Trophic Network of Fish Community

Dominant trophic groups differ in different fishing grounds (Table 6). Figures 35–42 in appendix 3a illustrate the trophic composition per *I Qoliqoli*. In summary the dominant trophic group in the big islands such as Viti Levu, Vanua Levu and Kadavu are herbivorous feeders. However, the dominant trophic group in Ovalau (big island) and Vusasivo (Vanua Levu) were invertebrate and piscivorous feeders. Similar results were observed in Ba and Rakiraki with the dominant of planktivorous feeder. High biomass of omnivorous and detritivorous was observed in Galoa.

Table 6: Dominant trophic network in each fish ground. The graph is on Figure 35 – Figure 42 in appendix 3a, which shows the biomass.

<b>I Qoliqoli</b>	<b>Dominant Trophic Group</b>
Nayau	Invertebrate Feeder
Oneata	Herbivore
Tikina Levuka	Invertebrate Feeder
Tikina Nasinu	Invertebrate and Piscivore Feeder
Naikorokoro	Herbivore
Namuana	Herbivore
Karoko	Herbivore
Vusasivo	Invertebrate and Piscivore
Rakiraki	Planktivore
Wananavu	Herbivore
Galoa	Omnivore and Detrivore
Namaqumaqua	Herbivore
Ba	Planktivore
Tavua	Herbivore
Lami	Herbivore
Mau	Herbivore

#### 4 DISCUSSION

Generally, the small bodied fishes such as Damselfishes (Pomacentridae) and Butterflyfishes (Chaetodontidae) are found to be in large numbers, while the bigger fishes (top predators and most important food fishes) are less abundant. The bigger fishes are the piscivorous/invertebrate feeders that are favoured by both artisanal and subsistence fishers. These are mostly, Snappers (Lutjanidae), Emperors (Lethrinidae), Groupers (Serranidae), Trevallys (Carangidae), Barracudas (Sphyraenidae), and Tunas (Scombridae). The islands *I Qoliqoli* (outside the main island, Viti Levu and Vanua Levu) had higher mean abundance and biomass of these fishes than the two major islands (Table 8 and 9). Studied by Jennings *et al.* (1995) in six *I Qoliqoli* in Fiji showed that the biomass of invertebrate feeders is significantly higher in the least intensively fished *I Qoliqoli*.

Habitat characteristics influence fish communities' structure (Jennings *et al.* 1997) or the distribution of fish within an *I Qoliqoli*. High mean abundance was seen in lagoon areas but low mean biomass (kg/transect), while high mean biomass (kg/transect) was detected in fore reef with less mean abundance (Figure 4 – 5). Fishes that were seen dominating the lagoon are mostly the non-food fish such as damselfish, butterflyfish and juveniles of important food fish. Lagoon provides good nursery ground for juvenile fishes because of its calm waters (less predation). Most of these fishes were found in small schools to protect them from predators.

Nearly all habitats showed high variance in mean length, which is an indication of some bigger fishes (such as top predators), sighted with a high abundance of small fish population in lagoon and back reef area. These top predators (Sharks, Snappers, Emperors, Barracudas, Trevallys, Tunas), were not sighted in all fishing grounds (within 500m<sup>2</sup> or transect) (Table 8), considering the widespread foraging behaviour of Snappers (Lutjanidae) (España 2003) as well as Sharks (Carchahinidae), Emperors (Lethrinidae), Barracudas and Tuna (Scombridae), Trevally (Carangidae).

The food fishes, Scaridae (Parrotfish), Acanthuridae (Surgeonfish), Mullidae (Goatfish) and Labridae (Wrasses) are found to be abundant in all fishing grounds (Table 5 and Table 6). However, these fishes are classified as minor commercial and are mostly used as subsistence, except *Bolbometapon muricatum* (Parrotfish) and *Cheilinus undulatus* (Wrasses), which are highly commercial. These fishes (Parrotfishes, Surgeonfishes, Wrasses and Goatfishes) are mainly herbivorous, omnivorous and detritivorous feeders (Table 9). Considering the feeding structure, high mean abundance of Acanthuridae and Scaridae were seen in the lagoon and reef area (Table 4). According to local fishermen in Viti Levu (especially in South Viti Levu), the dominant daily catch compositions are mainly the minor commercial fishes (Holocentridae, Mullidae, Scaridae, Labridae, Acanthuridae and Lethrinidae (*Lethrinus harak*). The removal of herbivorous could lead to ecological transition (McClanahan *et al.* 2002).

Higher mean abundance and biomass was sighted in Lagoon and Reef area of the outer islands than the main islands, for example, Nayau, Oneata of East Fiji, Naikorokoro, and Namuana of South Fiji (Figure 19 – 20). This is probably due to the findings by Kuster *et al.* (2005) in Lau Islands that the increase in outboard vessels appeared to have a social impact causing traditional shore based fishing activities of women to be less. During low tides women wade in this area to fish and glean for edible invertebrates such as bivalves, univalves, octopus and sea cucumbers. Now, men are taking the responsibility by spear dive in fore reefs and outer reefs.

Presumably, high population coupled with land effluents may cause the low mean abundance and biomass in lagoon and reef area of the main islands. Therefore, the habitat could be destroyed which may lead to fish moving to other undisturbed areas. This also influences the distribution of the community trophic group. The dominant fish group in the main islands of Viti Levu, Vanua Levu and Kadavu, are the herbivorous feeders (Table 6). Perhaps because of more development in these areas coupled with the annual rainfall of 2000 mm – 3000 mm, more land effluents enters the sea. This could lead to algal blooming, which could increase the abundance of herbivorous feeder. Algal bloom is common along the reef area on the Coral Coast (South Viti Levu), therefore overfishing of herbivorous fish could be a contributing factor whereby less herbivorous to feed on the algae.

Lami (East Viti Levu) fore reef has high fish abundance but low biomass compared to Nayau Island (East Fiji) which has low fish abundance but high biomass within the 10 m – 20 m depth range. The possible explanation for this is the low abundance of bigger fishes in the main island Viti Levu with the high abundance of small sizes between 10 m – 25 m depth. In Viti Levu, fishermen spend 3 -4 days out in the sea to meet the market demand. Marine protected area (MPA) could also affect the fish community in a fishing ground. In Tikina Levuka and Tikina Nasinu, certain areas of the fishing ground were closed during the survey ( 6 months before the survey were conducted).The intention was to breed and conserve the fish for the 2007 Annual Methodist Conference Meeting in the island of Ovalau. Therefore, the dominant trophic group sighted in the fishing grounds were the invertebrate feeder and piscivore feeder. According to Hoffmann (2002), reefs surrounding Ovalau and harbour surrounding Levuka have been affected by the PAFCO effluent. During the survey, villagers were complaining on the high number of sharks sighted around Levuka harbour. It could have been the herbivorous feeder to be the dominated group because the villagers depend on the sea for livelihood that is the higher removal of target fishes.

#### 4.1 Catch information

There was no catch data available to calculate the total yield of the fishing grounds or the exploitation of the stocks. Consequently, this study will improve the sampling scheme of the department and to impose the gathering of catch data/CPUE during the visual transect survey in fishing communities.

It should be considered that visual transect underestimated the abundance of cryptic fish species. Jennings and Polunin (1995) tested visual transect to estimate the biomass of *Lethrinus spp* in six *I Qoliqoli* in Fiji. The research discovered that the stock was not reflected in catches taken from the same reef areas at a similar time of day.

Therefore it must be noted that the study is to gather baseline biological information to allow DOF to understand the health and productivity of an *I Qoliqoli*, which then assist in the formulation of appropriate management plan for each *I Qoliqoli*.

### 5 CONCLUSION

Overall, fish abundance (density and biomass) is higher in outer islands than the main islands of Viti Levu and Vanua Levu. In all areas, there is a low fish count of the most important food fishes but high diversity. The dominant fishes in all fishing grounds are Damselfishes (Pomacentridae), Surgeonfishes (Acanthuridae), Parrotfishes (Scaridae), Wrasses (Labridae) and Goatfishes (Mullidae).

Lagoon showed high abundance but not much variation in the reef area, reef slope, back reef and fore reef. Fore reef had the highest biomass with higher variation. The lagoon of fishing grounds that is far from the high populated area (islands) had high biomass and abundance than the main island Viti Levu. The means size distribution pattern between habitats and depths in a fishing ground is rather consistent.

Herbivorous feeders is the dominant trophic group in the fishing grounds surveyed especially in high islands (Viti Levu, Vanua Levu and Kadavu), while invertebrate feeder is dominant in small islands. Highly commercial fishes (mostly invertebrate and piscivorous fish) are rare in areas with higher human population.

This study highlights the lack of basic catch data for the inshore fisheries in Fiji; therefore there is a gap in the sampling scheme. Catch data is needed for the determination of the status of the exploitation of stocks. This study will assist DOF to determine daily catch limits, gear restrictions and most of all the number of fishing licenses renting to artisanal fishers. Also the study enhances the analysis of biological data as well as data management.

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**APPENDIX****Appendix 1****Table 7: Total fish count and number of species per Family.**

<b>Family</b>	<b>Total Count of Fishes</b>	<b>No. Of Species</b>
Carangidae	63	10
Carcharhinidae	12	1
Haemulidae	26	7
Lethrinidae	752	17
Lutjanidae	2,842	22
Scombridae	363	4
Serranidae	216	21
Sphyraenidae	146	2
Scaridae	3,358	38
Acanthuridae	4,204	37
Labridae	1,401	67
Balistidae	249	12
Caesionidae	3,832	11
Gerreidae	21	
Hemiramphidae	95	2
Holocentridae	239	16
Mugilidae	65	3
Mullidae	1,121	15
Nemipteridae	370	8
Siganidae	1,057	14
Terapontidae	22	1
Zanclidae	139	1
Nemipteridae	370	8
Aulostomidae	3	1
Blennidae	76	7
Bothidae	1	1
Chaetodontidae	1,412	41
Chanidae	6	1
Cichlidae	2	1
Cirrhitidae	2	1
Dasyatidae	1	1
Diodontidae	6	2
Echeneidae	4	1
Eleotridae	2	1
Ephippidae	37	3
Fistularidae	2	1
Gerreidae	21	2
Gobiidae	27	8
Kyphosidae	47	3
Monacanthidae	6	5
Apogonidae	165	16
Ostraciidae	4	3
Pinguipedidae	91	6
Platycephalidae	4	1
Plotosidae	30	1
Polynemidae	5	1
Pomacanthidae	687	11
Pomacentridae	9,827	110
Priacanthidae	1	1
Scorpaenidae	3	1
Soleidae	7	1
Sparidae	2	1
Syngnathidae	5	1
Synodontidae	7	3
Muraenidae	2	2
Tetraodontidae	27	10
<b>TOTAL</b>	<b>33,094</b>	

## Appendix 2a

**Table 8: Abundance and Biomass per 500m<sup>2</sup> with standard error (SE) of the most important food fish families per I Qoliquoli. The list of families is arranged according to the high commercial to the least. However, Shark (Carcharhinidae) is placed on the list, because of the high commercial values of shark fins.**

Family	Nayau Is		Oneata Is		Naikorokoro		Namuana		Tikina Nasinu		Tikina Levuka		Karoko		Vusasivo	
	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass
Lutjanidae	54.3±55.1	28.1±30	27.5±48.4	3.6±4.6	3.6±4.6	0.6±0.6			1.6±3.6	1.3±2.8	25.5±34.6	29.7±54.8	3±4.2	1.2±1.7	3±4.2	1.2±1.7
Lethrinidae	8.4±11.2	2.4±4.1	7.5±15	0.1±0.3	4±3.4	0.9±1.4	4.3±4.3	0.7±0.7	9.4±2.7	2±0.9	3±2.8	3.1±3.3	1±1.4	0.1±0.2		
Serranidae	4.7±3	2.5±2.9	0.5±1	0.3±0.5	0.7±1.4	0±0.1	0.3±0.5	0±0.1	0.6±0.9	0.2±0.2	0.5±0.7	0.6±0.9	2±2.8	0.3±0.4	2±2.8	0.3±0.4
Carangidae	0.3±0.8	0.7±1.7					0.3±0.5	0.1±0.3	0.4±0.9	0.9±2.1			15±2.1			
Haemulidae	0.3±0.5	0.3±0.6														
Scombridae	0.3±0.5	1.3±2.3					16.5±21.8	3.9±4.8								
Sphyraenidae																
Scaridae	59.3±32.2	23.2±19.5	27.5±48.4	3.6±4.6	19.2±13.8	4.6±3.5	87.3±114.5	14.4±8.7	7.4±2.5	2.9±0.4	6±3.4	5±3.8	28±28.3	4.3±1.2	28±28.3	4.3±1.2
Acanthuridae	84.6±75.9	19.2±20.3	27.5±48.4	3.6±4.6	61.1±31.2	11.1±8.3	79.5±36.5	21.8±18.2	11.2±8.5	1.9±2.2	21.5±6.4	7.3±1.9	38.5±31.8	8.2±1.8	38.5±31.8	8.2±1.8
Mullidae	28.9±38.2	17.1±32.1	1.5±1.7	0.4±0.4	5.9±4.8	0.9±0.8	9.8±13.3	2.7±4.5	15.8±16.8	4.5±4.6	3.5±0.7	1.4±0.8	10±9.9	0.8±0.2	10±9.9	0.8±0.2
Labridae	16±17.7	5.2±6.3	9±11.6	0.7±0.8	14.6±7.6	2.3±1.6	13±6.7	1.7±0.8	9.4±2.7	2±0.9	4	3.1±3.3	10.5±10.6	1±0.2	10.5±10.6	1±0.2
Holocentridae	7.6±13.1	1±1.3			0.9±1.5	0.4±1			1.8±1.8	1.1±1.2	1±1.4	0.3±0.4	1.5±2.1	0.5±0.7	1.5±2.1	0.5±0.7
Nemipteridae	1.1±1.7	0.1±0.1	3.8±6.8	0.1±0.1	39.2±104.9	23.2±67.6	4.8±4	0.9±1.2	3.8±4.4	1±1.4	0.5±0.7	0.1±0.1	1±1.4	0.2±0.3	1±1.4	0.2±0.3
Siganidae	1.6±2.3	0.2±0.4			11.2±7.8	2.4±3.4	21±23.8	2±2.6	1.2±1.3	0.2±0.2	4.5±0.7	0.9±0.2				
Caesionidae	27.9±50.2	5.5±10			1±3	0.1±0.2			4.8±6.7	0.2±0.3	15±21.2	0.5±0.7				
Balistidae	4.7±3.6	2±3.3	2±4		2.1±2.2	2.4±5.7	2.5±3	0.5±0.7	1.2±0.8	0.6±1.2	3	8.1±2	1	0.1±0.2	1	0.1±0.2
Carcharhinidae	0.4±0.8	2.8	5.1													

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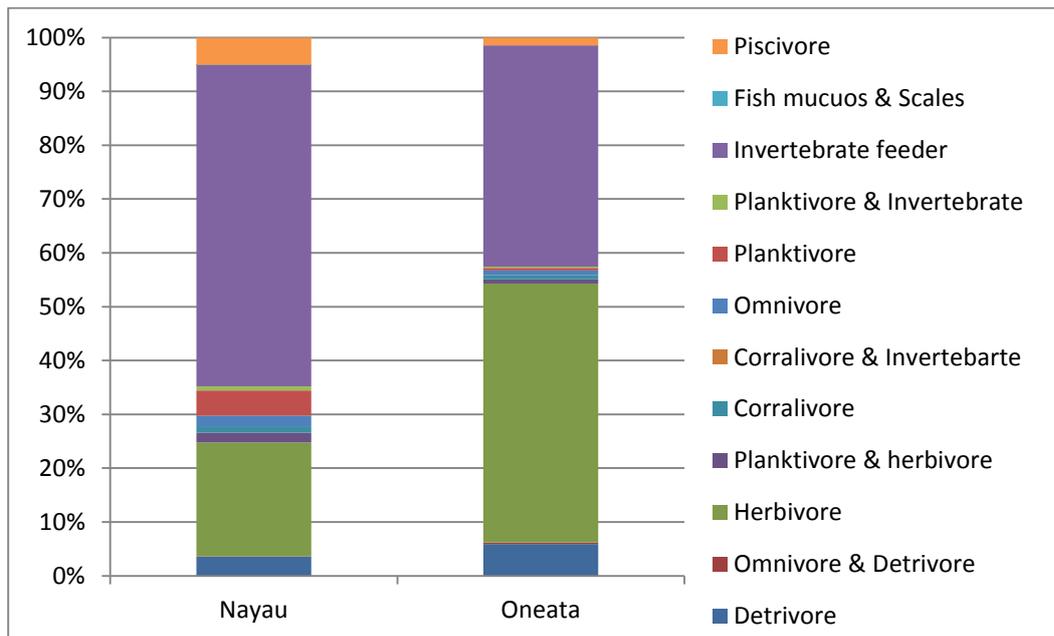
## Appendix 2b

**Table 9: Abundance and biomass per 500m<sup>2</sup> with standard error (SE) of the most important food fish families in the main island Viti Levu. The list of the species is arranged according to the high commercial to the least. However, Shark (Carcharhinidae) is placed on the list, because of its high value in shark fin trade.**

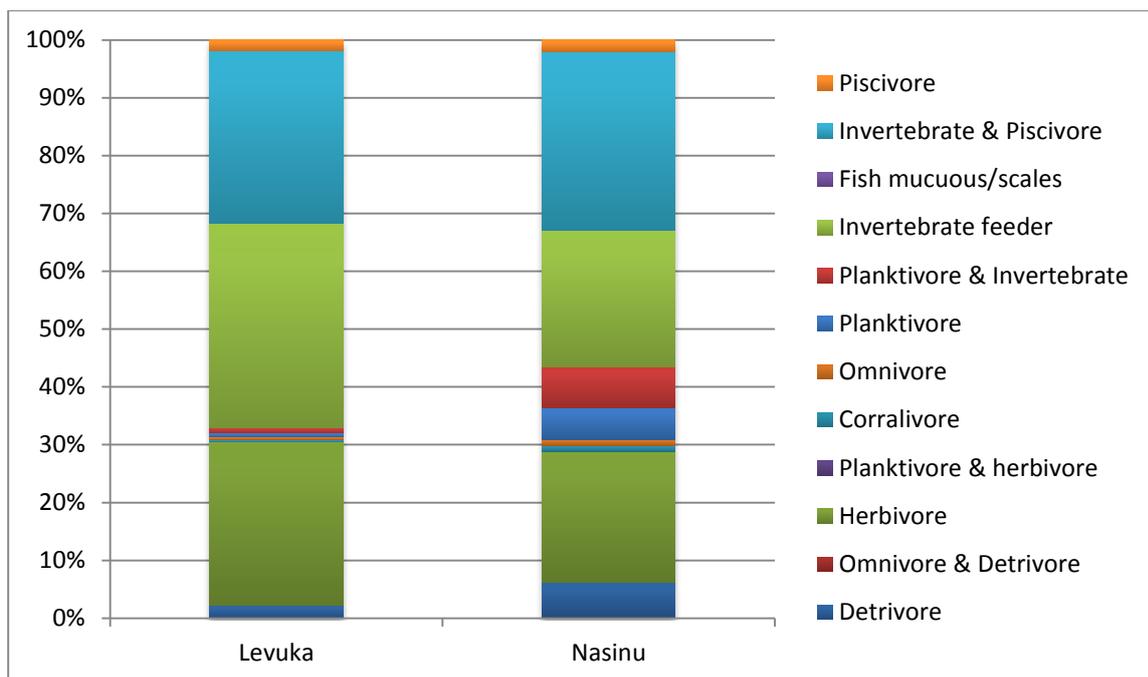
Family	Rakiraki		Wananavu		Ba		Tavua		Galoa		Namaqumaqua		Lami		Mau	
	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass	Density	Biomass
Lutjanidae	73±11.3	17.3±2.2	7.5±0.7	2.3±0.1	4.1±9.7	1.4±2.5	28.8±53.8	6.3±10.7	18.6±40.1	2.7±4.3			13±18.4	2.5±3.5	13±26.9	1.8±3.3
Lethrinidae			3±1.0	0.1±0.01			4.7±15.4	0.8±2.4	3.9±6.9	1.7±3.5	2.1±2.9	0.7±1.1			12.2±11.8	3.4±2.8
Serranidae			4.5±2.1	1±0	0.6±0.8	0.4±0.7	0.4±0.5	0.3±1	0.3±0.7	0.1±0.2			26.5±31.8	2.4±1.2	2.8±2.4	0.4±0.3
Carangidae			7.5±10.6	3±4.3			1±4	0.3±1.1	0.1±0.3	0.2±0.5	0.1±0.4	0.1±0.3				
Haemulidae	0.5±0.7	0.8±0.3			0.1±0.4		0.4±0.8	0.1±0.01								
Scombridae	50±70.7	24.4±34.5					0.3±0.7	0.2±0.6	0.2±1	0.1±0.3			32.5±46	13.9±19.7		
Sphyraenidae			5±7.1	2.2±3.1			2.3±9	0.3±1								
Scaridae	7±7.1	1.2±1.5	11±15.6	4±5.6	27.7±16	5.4±3.9	22.9±16.2	8.8±6.9	6±7	2.1±2.7	2.6±4.5	0.8±1.5	5.5±7.8	1.5±2.1	35.4±36.6	11.6±21
Acanthuridae	5.5±6.4	1.6±1.0	15.5±12	5.2±5.2	4±3.6	0.8±0.8	9.8±13.1	4±5.9	8.4±9.1	1.7±2.1	10±6.2	2.1±1.2	58.5±82.7	23.7±33.5	29±19.3	3.6±2.5
Mullidae	0.5±0.7	0.3±0.4	19±5.7	5.1±3.8	1.3±3	0.2±0.4	6±11	3.4±9.5	4.5±7	0.9±1.7	3.1±3.4	0.7±0.6	31±33.9	5.6±1.6	8±4.6	1.3±1.4
Labridae	4.5±2.1	0.3±0.1	4.5±6.4	0.6±0.9	10±6.7	1±0.9	9.1±4.7	1.3±1.2	3.9±5.9	0.4±0.8	5.9±4.2	0.9±0.9	19±26.9	0.7±1	6.2±10	0.3±0.4
Holocentridae			7±1.0	1.0±0.1	1.9±4.9	1.2±3.2	0.4±1	0.1±0.3	1.1±5.1	0.4±1.9	0.1±0.4					
Nemipteridae	3±1.4	0.2±0.1	2±0.1	0.4±0.1	6.3±6.2	0.5±0.4	4±2.2	0.4±0.3	0.7±1.6	0.1±0.4	0.8±0.9	0.1±0.1			1.6±1.7	0.2±0.2
Siganidae	1.5±0.7	0.3±0.1	17±5.7	2.4±2.2	3.4±2.9	0.3±0.2	18.9±50.4	9.4±35.3	0.6±1.6	0±0.1	1.4±2	0.2±0.3	62±87.7	2.3±3.3	8.4±15.6	0.8±1.3
Caesionidae	81.5±54.4	14.9±3.6	10±14.1	0.3±0.4	112.1±141	8.5±11.6	31.4±87.3	3.7±10.6	20±38.1	1.5±3						
Balistidae	0.5±0.7		1.5±0.7	0.3±0.3			1.3±2	0.4±1	0.8±1.8	0.1±0.2	2±2.8	0.4±0.7	0.5±0.7	0±0.1	0.8±1.8	
Carcharhinidae							0.2±0.4	1.5±4.3							0.2±0.4	0.9±2
Mugilidae			12.5±17.7	16.1±8.7												

Gerreidae			3.5±0.7	0.6±0.1												
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### Appendix 3a



**Figure 35: Trophic network of fish community in Nayau Island and Oneata Island (Lau Group), East Fiji.**



**Figure 36: Trophic network of fish community in Tikina Levuka and Tikina Nasinu, Ovalau, Central Fiji.**

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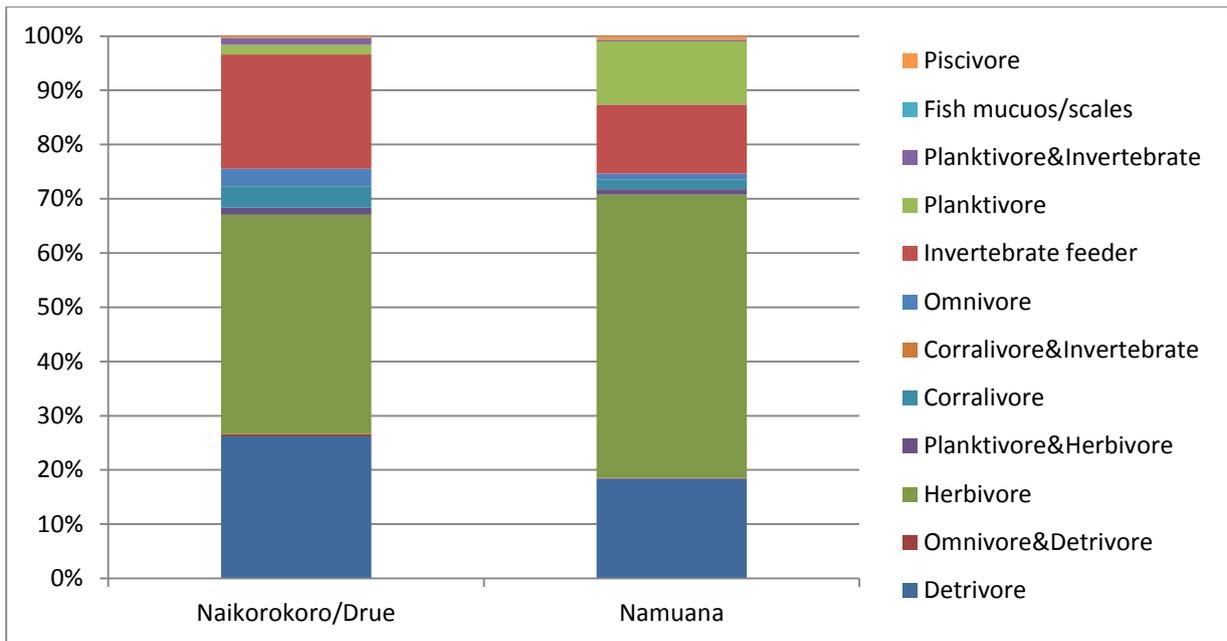


Figure 37: Trophic network of fish community in Naikorokoro and Namuana, Kadavu Island, South Fiji.

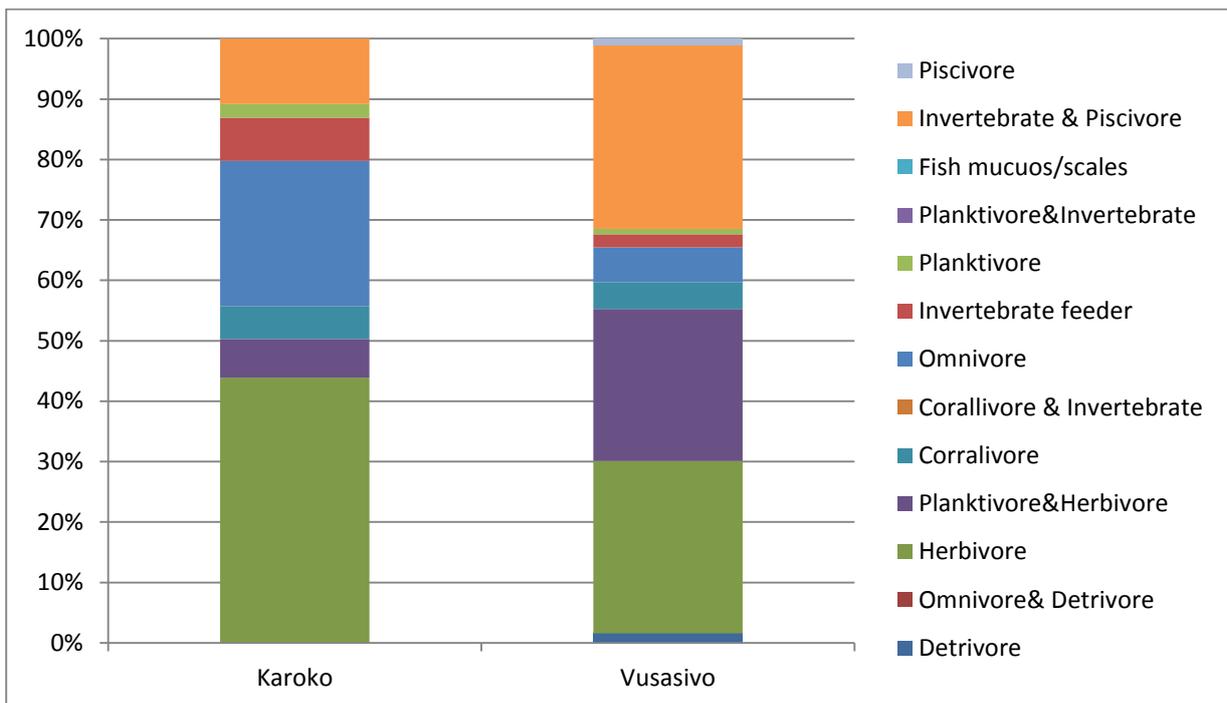


Figure 38: Trophic network of fish community in Karoko (Tunuloa) and Vusasivo (Natewa), North Fiji.

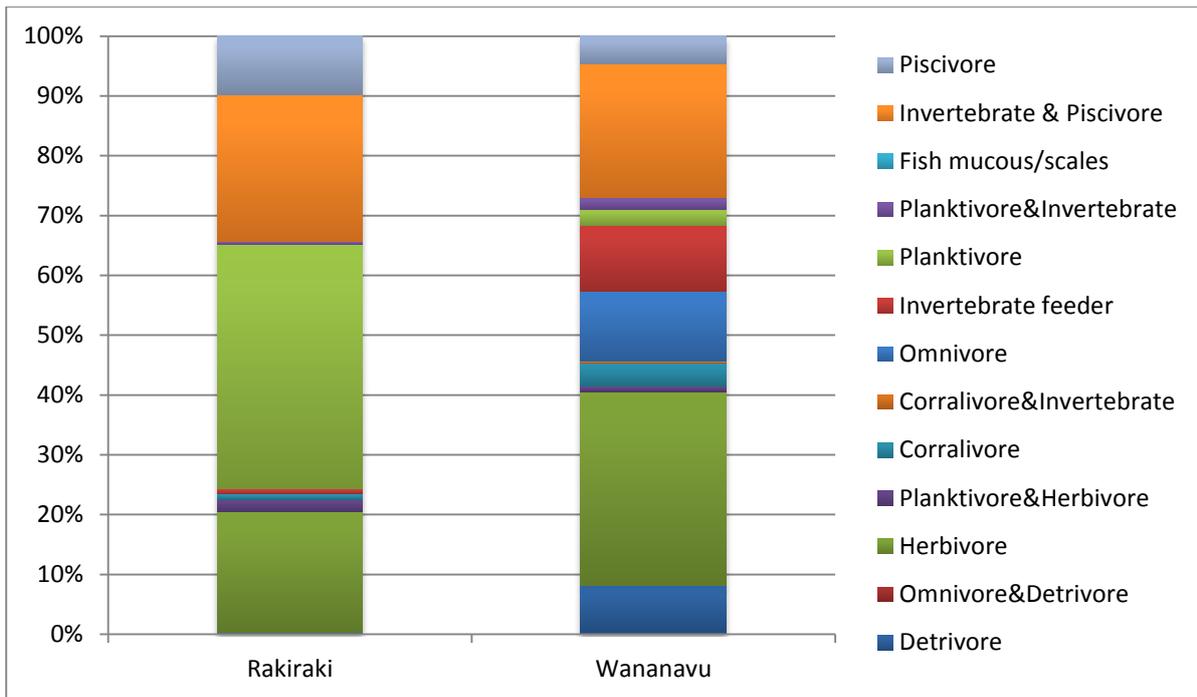


Figure 39: Trophic network of fish community in Rakiraki and Wananavu, Ra, North Viti Levu.

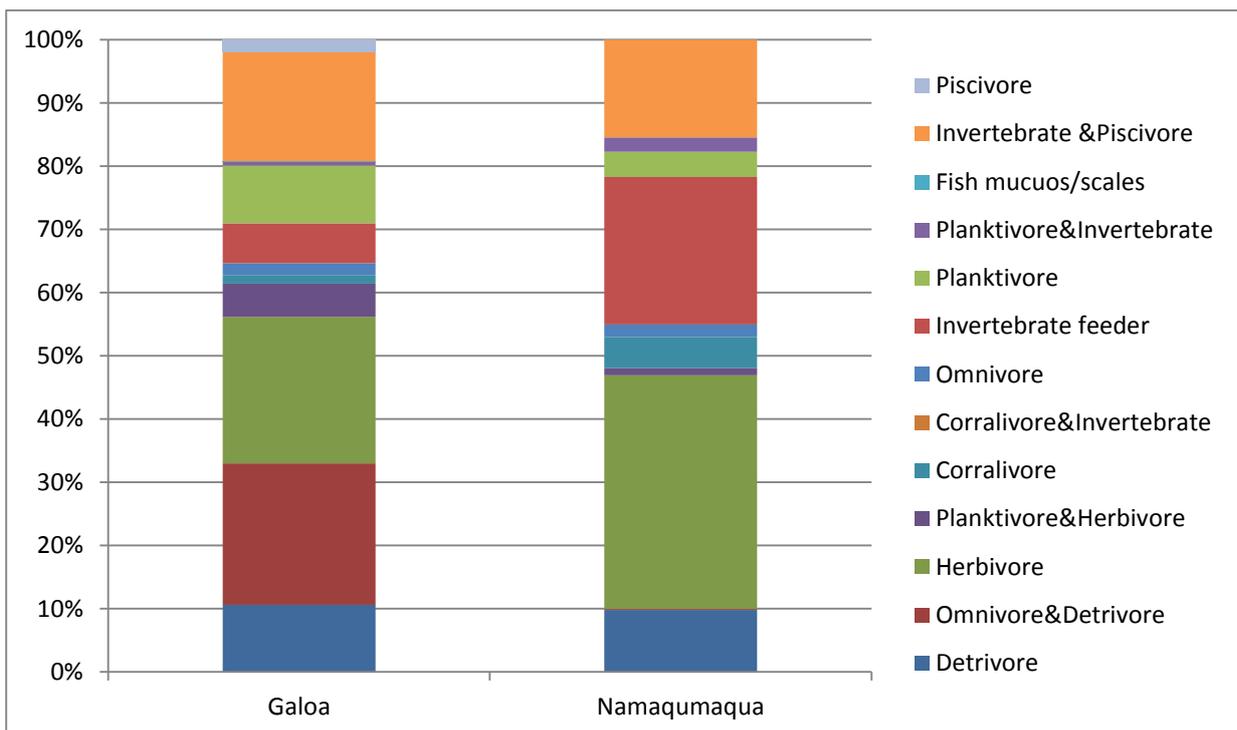


Figure 40: Trophic network of fish community in Galoa and Namaqumaqua, Serua, South Viti Levu.

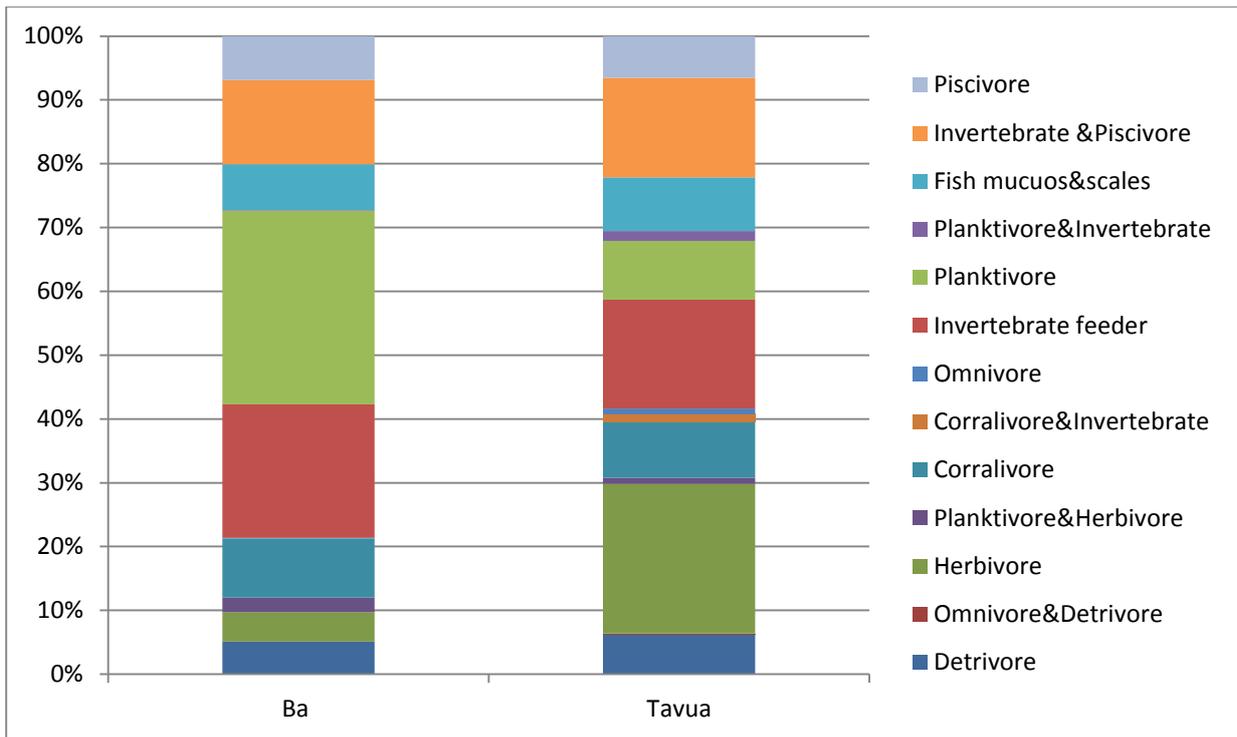


Figure 41: Trophic network of fish community in Ba and Tavua, Ba Province, West Viti Levu.

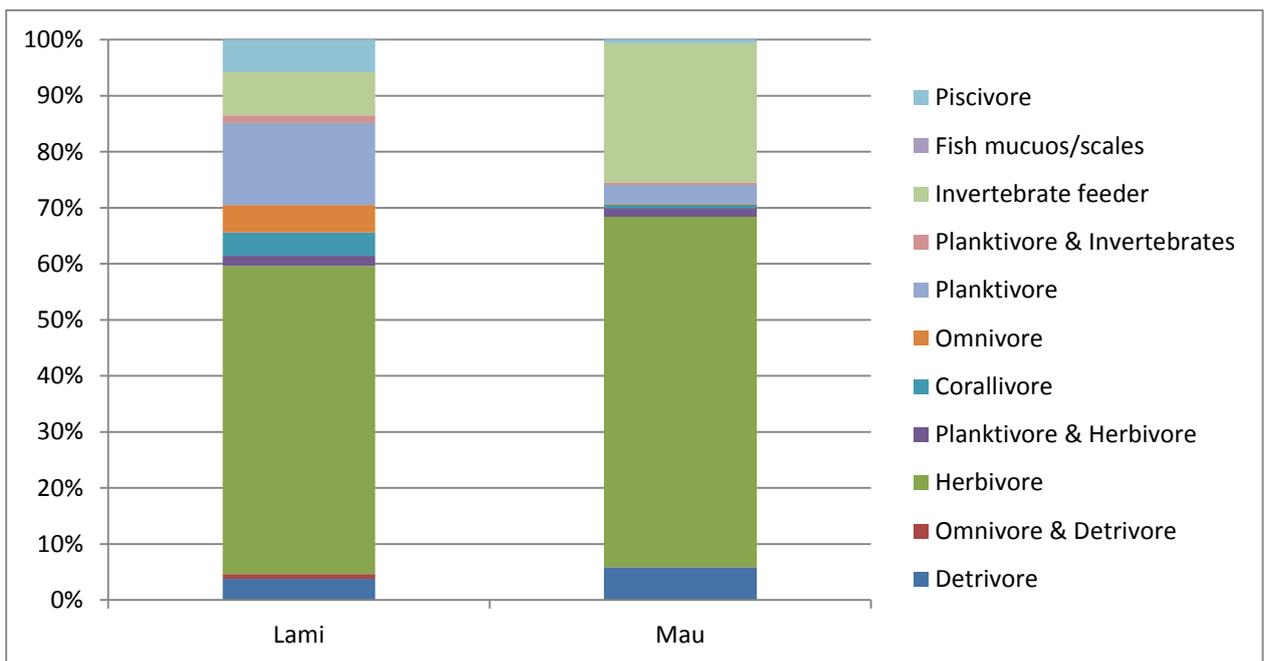


Figure 42: Trophic network of fish community in Lami and Mau, East Viti Levu.

**Appendix 3b****Table 10: Fish species assigned into trophic groups (Jennings et al. 1995 and Fish Base (www.fishbase.org)).**

<b>Family</b>	<b>Trophic Group</b>
<b>Acanthuridae</b>	
Acanthurus grammoptilus	Herbivorous
Acanthurus maculiceps	Herbivorous
Acanthurus nigrofuscus	Herbivorous
Acanthurus achilles	Herbivorous
Acanthurus albipectoralis	Herbivorous
Acanthurus auranticavus	Herbivorous
Acanthurus blochii	Herbivorous
Acanthurus fowleri	Herbivorous
Acanthurus grandoculis	Herbivorous
Acanthurus guttatus	Herbivorous
Acanthurus japonicus	Herbivorous
Acanthurus leuchocheilus	Herbivorous
Acanthurus lineatus	Herbivorous
Acanthurus mata	Herbivorous
Acanthurus nigricans	Herbivorous
Acanthurus nigricauda	Herbivorous
Acanthurus nigroris	Herbivorous
Acanthurus nubilus	Herbivorous
Acanthurus olivaceous	Herbivorous
Acanthurus pyroferus	Herbivorous
Acanthurus quttus	Herbivorous
Acanthurus sp.	Herbivorous
Acanthurus thompsoni	Herbivorous
Acanthurus triostegus	Herbivorous
Acanthurus xanthopterus	Herbivorous
Ctenochaetus binotatus	Herbivorous
Ctenochaetus citrinellus	Herbivorous
Ctenochaetus cyanocheilus	Herbivorous
Ctenochaetus marginatus	Herbivorous
Ctenochaetus sp.	Herbivorous
Ctenochaetus striatus	Herbivorous
Naso annulatus	Herbivorous
Naso brachycentron	Herbivorous
Naso brevirostis	Herbivorous
Naso caesius	Herbivorous
Naso hexacanthus	Herbivorous
Naso lituratus	Herbivorous
Naso unicornis	Herbivorous
Naso vlamingii	Herbivorous
Zebrasoma desjardini	Herbivorous

<i>Zebrasoma rostratum</i>	Herbivorous
<i>Zebrasoma scopas</i>	Herbivorous
<i>Zebrasoma veliferum</i>	Herbivorous
<b>Apogonidae</b>	
<i>Apogon compressus</i>	Planktivore & Invertebrate
<i>Apogon cookii</i>	Planktivore & Invertebrate
<i>Apogon dispar</i>	Planktivore & Invertebrate
<i>Apogon guamensis</i>	Planktivore & Invertebrate
<i>Apogon heptacanthus</i>	Planktivore & Invertebrate
<i>Apogon nigrofasciatus</i>	Planktivore & Invertebrate
<i>Apogon novemfasciatus</i>	Planktivore & Invertebrate
<i>Apogon novemferum</i>	Planktivore & Invertebrate
<i>Archamia zosterophora</i>	Planktivore & Invertebrate
<i>Cheilodipterus quinquelineatus</i>	Invertebrate & Piscivore
<i>Cheilodipterus sp.</i>	Invertebrate & Piscivore
<i>Cheilodipterus macrodon</i>	Piscivore
<i>Fowleria variegata</i>	Planktivore & Invertebrate
<i>Sphaeramia orbicularis</i>	Invertebrate feeder
<b>Balistidae</b>	
<i>Balistapus undulatus</i>	Omnivore
<i>Rhinecanthus aculeatus</i>	Omnivore
<i>Rhinecanthus lunula</i>	Omnivore
<i>Rhinecanthus rectangulus</i>	Omnivore
<i>Rhinecanthus verrucosus</i>	Omnivore
<i>Sufflamen bursa</i>	Invertebrate feeder
<i>Sufflamen chrysopterus</i>	Invertebrate feeder
<i>Xanthichthys auromarginatus</i>	Planktivore.
<i>Pseudobalistes flavimarginatus</i>	Invertebrate feeder
<i>Melichthys vidua</i>	Omnivore
<i>Abalistes stellatus</i>	Invertebrate feeder
<b>Bleniidae</b>	
<i>Cirripectes chelomatus</i>	Herbivore
<i>Glyptoparus delicatulus</i>	Herbivore
<i>Plagiotremus landandus</i>	Fish mucuous & scales
<i>Plagiotremus rhinorhynchus</i>	Fish mucuous & scales
<i>Plagiotremus sp.</i>	Fish mucuous & scales
<i>Salarias fasciatus</i>	Herbivore
<i>Meiacanthus atrodorsalis</i>	Planktivore & Invertebrate
<b>Caesionidae</b>	
<i>Caesio caerulea</i>	Planktivore
<i>Caesio lunaris</i>	Planktivore
<i>Caesio teres</i>	Planktivore
<i>Caesio varilineata</i>	Planktivore
<i>Caesio xanthorota</i>	Planktivore

Pterocaesio digramma	Planktivore
Pterocaesio marri	Planktivore
Pterocaesio pisang	Planktivore
Pterocaesio tile	Planktivore
Pterocaesio trilineata	Planktivore
Gymnocaesio gymnoptera	Planktivore
<b>Carangidae</b>	
Caranx ignobilis	Invertebrate feeder & Piscivore
Caranx melampygus	Invertebrate feeder & Piscivore
Gnathanodon speciosus	Invertebrate feeder & Piscivore
Scomberoides commersonianus	Invertebrate feeder & Piscivore
Scomberoides lysan	Invertebrate feeder & Piscivore
Elagatis bipinnulata	Invertebrate feeder & Piscivore
Caranx lugubris	Piscivore
Caranx papuensis	Piscivore
Carangoides plagiotaenia	Planktivore & Invertebrate
Carangoides sp.	Planktivore & Invertebrate
<b>Carchahinidae</b>	
Triaenodon obesus	Invertebrate feeder & Piscivore
<b>Chanidae</b>	
Chanos chanos	Herbivore
<b>Chichlidae</b>	
Hemichromis sp.	Invertebrate feeder & Piscivore
<b>Cirrhitidae</b>	
Paracirrhites forsteri	Invertebrate feeder & Piscivore
<b>Chaetodontidae</b>	
Chaetodon auriga	Corralivore
Chaetodon austriacus	Corralivore
Chaetodon baronessa	Corralivore
Chaetodon benetti	Corralivore
Chaetodon cellicaudus	Corralivore
Chaetodon citrinellus	Corralivore
Chaetodon decussatus	Corralivore
Chaetodon ephippium	Corralivore
Chaetodon falcula	Corralivore
Chaetodon fasciatus	Corralivore
Chaetodon guentheri	Corralivore
Chaetodon kleinii	Corralivore
Chaetodon lineolatus	Corralivore
Chaetodon lunula	Corralivore
Chaetodon lunulatus	Corralivore
Chaetodon melannotus	Corralivore
Chaetodon mertensii	Corralivore
Chaetodon mesoleucus	Corralivore

Chaetodon meyersi	Corralivore
Chaetodon ocellicaudus	Corralivore
Chaetodon ornatissimus	Corralivore
Chaetodon ornatus	Corralivore
Chaetodon oxycephalus	Corralivore
Chaetodon pelewensis	Corralivore
Chaetodon plebeius	Corralivore
Chaetodon punctatofasciatus	Corralivore
Chaetodon rafflesi	Corralivore
Chaetodon reticulatus	Corralivore
Chaetodon semeion	Corralivore
Chaetodon sp.	Corralivore
Chaetodon speculum	Corralivore
Chaetodon trifascialis	Corralivore
Chaetodon trifasciatus	Corralivore
Chaetodon ulientensis	Corralivore
Chaetodon unimaculatus	Corralivore
Chaetodon vagabundus	Corralivore
<b>Dasyatidae</b>	
Taeniura lymma	Invertebrate feeder
<b>Diodontidae</b>	
Diodon hystrix	Invertebrate feeder
Diodon liturosus	Invertebrate feeder
<b>Echeneidae</b>	
Echeneis naucrates	Piscivore
<b>Eleotridae</b>	
Oxyeleotris sp.	Invertebrate feeder & Piscivore
<b>Ephippidae</b>	
Platax boersii	Omnivore
Platax orbicularis	Omnivore
Platax teira	Omnivore
<b>Fistularidae</b>	
Fistularia commersonii	Invertebrate feeder & Piscivore
<b>Gerreidae</b>	
Gerres argyreus	Detrivore
Gerres oyena	Planktivore & Invertebrate
<b>Gobiidae</b>	
Amblyeleotris randalli	Planktivore
Amblygobius nocturnus	Omnivore & Detrivore
Amblygobius phalaena	Omnivore & Detrivore
Cryptocentrio cyanotaenia	Planktivore
Istigobius decoratus	Herbivore
Stonogobiops xanthorhinica	Planktivore
Valenciennesa sexguttata	Invertebrate feeder

Valencienna strigata	Invertebrate feeder
<b>Haemulidae</b>	
Haemulon sp.	Omnivore
Plectorhinchus albovittatus	Invertebrate feeder
Plectorhinchus chaetodonoides	Invertebrate feeder
Plectorhinchus multivittatum	Invertebrate feeder
Plectorhinchus pictus	Invertebrate feeder
Plectorhinchus sp.	Invertebrate feeder
Plectorhinchus vittatus	Invertebrate feeder
<b>Hemiramphidae</b>	
Hemiramphus far	Herbivore
Hyporhamphus dussumieri	Planktivore
<b>Holocentridae</b>	
Myripristis adusta	Planktivore
Myripristis amaena	Invertebrate feeder
Myripristis hexagon	Planktivore
Myripristis murdjan	Planktivore
Myripristis pralinia	Planktivore
Myripristis violacea	Invertebrate feeder
Myripristis vittata	Planktivore
Neoniphon argenteus	Invertebrate feeder
Neoniphon opercularis	Invertebrate feeder
Neoniphon sammara	Invertebrate feeder
Sargocentron candimaculatum	Invertebrate feeder
Sargocentron diadema	Planktivore
Sargocentron ittodai	Invertebrate feeder
Sargocentron spiniferum	Invertebrate feeder & Piscivore
Sargocentron tiere	Invertebrate feeder & Piscivore
Sargocentron violaceum	Invertebrate feeder
<b>Kyphosidae</b>	
Kyphosus bigibbus	Herbivore
Kyphosus cinerascens	Herbivore
Kyphosus vaigiensis	Invertebrate feeder
<b>Labridae</b>	
Anampes geographicus	Invertebrate feeder
Anampses caeruleopunctatus	Invertebrate feeder
Anampses geographicus	Invertebrate feeder
Anampses meleagrides	Invertebrate feeder
Anampses neoguinaicus	Invertebrate feeder
Bodianus bilunulatus	Invertebrate feeder
Bodianus loxozonus	Invertebrate feeder
Bodianus mesothorax	Invertebrate feeder
Cheilinus chlorourus	Invertebrate feeder
Cheilinus digrammus	Piscivore

<i>Cheilinus fasciatus</i>	Invertebrate feeder
<i>Cheilinus lunulatus</i>	Invertebrate feeder
<i>Cheilinus oxycephalus</i>	Invertebrate feeder
<i>Cheilinus trilobatus</i>	Invertebrate feeder
<i>Cheilinus undulatus</i>	Invertebrate feeder
<i>Cheilio inermis</i>	Invertebrate feeder
<i>Coris aygula</i>	Invertebrate feeder
<i>Coris bulbifrons</i>	Invertebrate feeder
<i>Coris gaimard</i>	Invertebrate feeder
<i>Epibulus epibulus</i>	Invertebrate & Piscivore
<i>Epibulus insidiator</i>	Invertebrate & Piscivore
<i>Gomphosus varius</i>	Invertebrate feeder
<i>Halichoeres hortulanus</i>	Invertebrate feeder
<i>Halichoeres argus</i>	Invertebrate feeder
<i>Halichoeres binotopsis</i>	Invertebrate feeder
<i>Halichoeres biocellatus</i>	Invertebrate feeder
<i>Halichoeres chloropterus</i>	Invertebrate feeder
<i>Halichoeres dussumieri</i>	Invertebrate feeder
<i>Halichoeres hortulanus</i>	Invertebrate feeder
<i>Halichoeres leucurus</i>	Invertebrate feeder
<i>Halichoeres margaritaceus</i>	Invertebrate feeder
<i>Halichoeres marginatus</i>	Invertebrate feeder
<i>Halichoeres melanurus</i>	Invertebrate feeder
<i>Halichoeres nigrescens</i>	Invertebrate feeder
<i>Halichoeres ornatissimus</i>	Invertebrate feeder
<i>Halichoeres prosopeion</i>	Invertebrate feeder
<i>Halichoeres sp.</i>	Invertebrate feeder
<i>Halichoeres trimaculatus</i>	Invertebrate feeder
<i>Halichoeres vrolikii</i>	Invertebrate feeder
<i>Hemigymnus fasciatus</i>	Invertebrate feeder & Piscivore
<i>Hemigymnus melapterus</i>	Invertebrate feeder & Piscivore
<i>Hologymnosus doliatus</i>	Invertebrate feeder & Piscivore
<i>Labroides bicolor</i>	Invertebrate feeder
<i>Labroides dimidiatus</i>	Invertebrate feeder
<i>Labroides pectoralis</i>	Fish mucous & scales
<i>Labropsis australis</i>	Corralivore
<i>Novaculichthys taeniourus</i>	Invertebrate feeder
<i>Oxycheilinus digrammus</i>	Invertebrate feeder
<i>Oxycheilinus rhodochrous</i>	Invertebrate feeder
<i>Oxycheilinus sp.</i>	Invertebrate feeder
<i>Oxycheilinus unifasciatus</i>	Invertebrate feeder & Piscivore
<i>Pseudodax mollucanus</i>	Omnivore
<i>Pseudojuloides cerasinus</i>	Invertebrate feeder
<i>Stethojulis bandanensis</i>	Planktivore & Invertebrate

<i>Stethojulis notialis</i>	Invertebrate feeder
<i>Stethojulis strigiventer</i>	Detrivore
<i>Thalassoma amblycephalum</i>	Planktivore.
<i>Thalassoma hardwicke</i>	Invertebrate feeder
<i>Thalassoma intesceris</i>	Invertebrate feeder
<i>Thalassoma janseni</i>	Invertebrate feeder
<i>Thalassoma klunzingeri</i>	Invertebrate feeder
<i>Thalassoma lunare</i>	Invertebrate feeder
<i>Thalassoma lutescens</i>	Invertebrate feeder
<i>Thalassoma purpureum</i>	Invertebrate feeder
<i>Thalassoma quinquevittatum</i>	Invertebrate feeder
<i>Thalassoma sp.</i>	Invertebrate feeder
<i>Thalassoma trilobatum</i>	Invertebrate feeder
<b>Lethrinidae</b>	
<i>Gnathodentex aureolineatus</i>	Invertebrate feeder & Piscivore
<i>Gymnocranius frenatus</i>	Invertebrate feeder.
<i>Gymnocranius grandoculis</i>	Invertebrate feeder & Piscivore
<i>Gymnocranius microdon</i>	Invertebrate feeder
<i>Lethrinus atkinsoni</i>	Invertebrate feeder & Piscivore
<i>Lethrinus erythracanthus</i>	Invertebrate feeder
<i>Lethrinus harak</i>	Invertebrate feeder & Piscivore
<i>Lethrinus mahsena</i>	Invertebrate feeder & Piscivore
<i>Lethrinus microdon</i>	Invertebrate feeder & Piscivore
<i>Lethrinus nebulosus</i>	Invertebrate feeder & Piscivore
<i>Lethrinus obsoletus</i>	Invertebrate feeder & Piscivore
<i>Lethrinus olivaceus</i>	Invertebrate feeder & Piscivore
<i>Lethrinus ornatus</i>	Invertebrate feeder & Piscivore
<i>Lethrinus rubrioperculatus</i>	Invertebrate feeder & Piscivore
<i>Lethrinus sp.</i>	Invertebrate feeder & Piscivore
<i>Lethrinus xanthochilus</i>	Invertebrate feeder & Piscivore
<i>Monotaxis grandoculis</i>	Invertebrate feeder
<b>Lutjanidae</b>	
<i>Aphareus furca</i>	Invertebrate feeder & Piscivore
<i>Aprion virescens</i>	Piscivore
<i>Lutjanus argentimaculatus</i>	Piscivore
<i>Lutjanus biguttatus</i>	Invertebrate feeder & Piscivore
<i>Lutjanus bohar</i>	Piscivore
<i>Lutjanus bouton</i>	Invertebrate feeder & Piscivore
<i>Lutjanus ehrenbergii</i>	Invertebrate feeder & Piscivore
<i>Lutjanus fulviflammus</i>	Invertebrate feeder & Piscivore
<i>Lutjanus fulvus</i>	Invertebrate feeder & Piscivore
<i>Lutjanus gibbus</i>	Invertebrate feeder & Piscivore
<i>Lutjanus kasmira</i>	Invertebrate feeder & Piscivore
<i>Lutjanus lemniscatus</i>	Invertebrate feeder & Piscivore

<i>Lutjanus maxweberi</i>	Invertebrate feeder & Piscivore
<i>Lutjanus monostigma</i>	Invertebrate feeder & Piscivore
<i>Lutjanus quinquelineatus</i>	Invertebrate feeder & Piscivore
<i>Lutjanus rivulatus</i>	Invertebrate feeder & Piscivore
<i>Lutjanus russelli</i>	Invertebrate feeder & Piscivore
<i>Lutjanus semicinctus</i>	Piscivore
<i>Lutjanus vittus</i>	Invertebrate feeder & Piscivore
<i>Macolor macularis</i>	Planktivore
<i>Macolor niger</i>	Planktivore
<i>Symphorichthys spilurus</i>	Invertebrate feeder & Piscivore
<b>Monacanthidae</b>	
<i>Aluterus scriptus</i>	Omnivore
<i>Amanses scopas</i>	Omnivore
<i>Oxymonacanthus longirostris</i>	Corralivore
<i>Paraluteres prionurus</i>	Invertebrate feeder
<i>Pseudalutarius</i> sp.	Invertebrate feeder
<b>Mugilidae</b>	
<i>Crenimugil crenilabis</i>	Omnivore
<i>Liza vaigiensis</i>	Omnivore
<i>Valamugi seheli</i>	Herbivore
<b>Mullidae</b>	
<i>Mulloidichthys flavolineatus</i>	Invertebrate feeder & Piscivore
<i>Mulloidichthys mimicus</i>	Invertebrate feeder & Piscivore
<i>Mulloidichthys pflugeri</i>	Invertebrate feeder & Piscivore
<i>Mulloidichthys vanicolensis</i>	Invertebrate feeder
<i>Parupeneus barberinoides</i>	Invertebrate feeder
<i>Parupeneus barberinus</i>	Planktivore & Invertebrate
<i>Parupeneus bifasciatus</i>	Invertebrate feeder & Piscivore
<i>Parupeneus ciliatus</i>	Invertebrate feeder & Piscivore
<i>Parupeneus cyclostomus</i>	Invertebrate feeder & Piscivore
<i>Parupeneus heptacanthus</i>	Invertebrate feeder & Piscivore
<i>Parupeneus indicus</i>	Invertebrate feeder & Piscivore
<i>Parupeneus multifasciatus</i>	Invertebrate feeder
<i>Parupeneus</i> sp.	Invertebrate feeder & Piscivore
<i>Parupeneus trifasciatus</i>	Invertebrate feeder & Piscivore
<i>Upeneus moleccensis</i>	Invertebrate feeder & Piscivore
<b>Muraenidae</b>	
<i>Gymnothorax javanicus</i>	Invertebrate feeder & Piscivore
<i>Gymnothorax richardsoni</i>	Invertebrate feeder & Piscivore
<b>Nemipteridae</b>	
<i>Scolopsis affinis</i>	Invertebrate feeder
<i>Scolopsis bilineata</i>	Invertebrate feeder & Piscivore
<i>Scolopsis frenatus</i>	Invertebrate feeder
<i>Scolopsis lineatus</i>	Invertebrate feeder & Piscivore

<i>Scolopsis monogramma</i>	Invertebrate feeder & Piscivore
<i>Scolopsis</i> sp.	Invertebrate feeder
<i>Scolopsis temporalis</i>	Planktivore & Invertebrate
<i>Scolopsis trilineatus</i>	Planktivore & Invertebrate
<b>Ostraciidae</b>	
<i>Ostracion cubicus</i>	Omnivore
<i>Ostracion meleagris</i>	Omnivore
<i>Rhynchostracion nasus</i>	Omnivore
<b>Pinguipedidae</b>	
<i>Parapercis clathrata</i>	Planktivore
<i>Parapercis cylindrica</i>	Planktivore
<i>Parapercis hexophthalma</i>	Planktivore
<i>Parapercis millepunctata</i>	Planktivore
<i>Synodus variegatus</i>	Piscivore
<b>Platycephalidae</b>	
<i>Papilloculiceps</i> sp.	Invertebrate feeder
<b>Plotosidae</b>	
<i>Plotosus lineatus</i>	Planktivore & Invertebrate
<b>Polynemidae</b>	
<i>Polydactylus sexfilis</i>	Invertebrate feeder
<b>Pomacanthidae</b>	
<i>Pomacanthus imperator</i>	Invertebrate feeder
<i>Pomacanthus semicirculatus</i>	Omnivore
<i>Pygoplites diacanthus</i>	Invertebrate feeder
<i>Centropyge bispinosus</i>	Herbivorous
<i>Centropyge centropyge</i>	Herbivorous
<i>Centropyge colini</i>	Herbivorous
<i>Centropyge flavissimus</i>	Herbivorous
<i>Centropyge heraldi</i>	Herbivorous
<i>Centropyge multicolor</i>	Herbivorous
<i>Centropyge multispinis</i>	Herbivorous
<i>Centropyge</i> sp.	Herbivorous
<i>Centropyge tibicen</i>	Herbivorous
<i>Centropyge woodheadi</i>	Herbivorous
<b>Pomacentridae</b>	
<i>Abudefduf bengalensis</i>	Omnivore
<i>Abudefduf sexfasciatus</i>	Planktivore & Herbivore
<i>Abudefduf sordidus</i>	Omnivore
<i>Abudefduf vaigiensis</i>	Planktivore & Herbivore
<i>Abudefduf whitleyi</i>	Planktivore
<i>Altrichthys azurelineatus</i>	Planktivore
<i>Amblyglyphidodon aureus</i>	Planktivore
<i>Amblyglyphidodon batunai</i>	Planktivore
<i>Amblyglyphidodon curacao</i>	Planktivore & Herbivore

<i>Amblyglyphidodon indicus</i>	Planktivore
<i>Amblyglyphidodon leucogaster</i>	Planktivore
<i>Amblyglyphidodon sp.</i>	Planktivore
<i>Amblyglyphidodon ternatensis</i>	Planktivore
<i>Amphiprion allardi</i>	Planktivore
<i>Amphiprion chrysopterus</i>	Planktivore
<i>Amphiprion clarkii</i>	Planktivore
<i>Amphiprion melanopus</i>	Planktivore
<i>Amphiprion rubrocinctus</i>	Planktivore
<i>Amphiprion sp.</i>	Planktivore
<i>Cheiloprion labiatus</i>	Corralivore
<i>Chromis agilis</i>	Planktivore & Herbivore
<i>Chromis analis</i>	Planktivore
<i>Chromis atripectoralis</i>	Planktivore
<i>Chromis caudalis</i>	Planktivore
<i>Chromis chrysur</i>	Planktivore
<i>Chromis dimidiata</i>	Planktivore & Herbivore
<i>Chromis elerae</i>	Planktivore & Herbivore
<i>Chromis iomelas</i>	Planktivore & Herbivore
<i>Chromis margaritifer</i>	Planktivore
<i>Chromis retrofasciata</i>	Planktivore & Herbivore
<i>Chromis sp.</i>	Planktivore & Herbivore
<i>Chromis ternatensis</i>	Planktivore & Herbivore
<i>Chromis viridis</i>	Planktivore.
<i>Chromis weberi</i>	Planktivore & Herbivore
<i>Chromis xanthura</i>	Planktivore & Herbivore
<i>Chrysiptera biocellata</i>	Herbivore
<i>Chrysiptera brownriggii</i>	Omnivore
<i>Chrysiptera caeruleolineata</i>	Invertebrate feeder
<i>Chrysiptera cyanea</i>	Omnivore
<i>Chrysiptera glauca</i>	Herbivore
<i>Chrysiptera parasema</i>	Planktivore
<i>Chrysiptera rollandi</i>	Planktivore
<i>Chrysiptera springeri</i>	Planktivore & Herbivore
<i>Chrysiptera starcki</i>	Planktivore & Herbivore
<i>Chrysiptera talboti</i>	Planktivore
<i>Chrysiptera taupou</i>	Planktivore & Herbivore
<i>Chrysiptera tricineta</i>	Planktivore & Herbivore
<i>Chrysiptera unimaculata</i>	Planktivore & Herbivore
<i>Dascyllus aruanus</i>	Omnivore & Detrivore
<i>Dascyllus flavicaudus</i>	Omnivore & Detrivore
<i>Dascyllus melanurus</i>	Omnivore & Detrivore
<i>Dascyllus reticulatus</i>	Omnivore & Detrivore
<i>Dascyllus trimaculatus</i>	Omnivore & Detrivore

<i>Dischistodus chrysopoecilus</i>	Herbivore
<i>Dischistodus prosopotaenia</i>	Herbivore
<i>Hemiglyphidodon plegiometopon</i>	Herbivore
<i>Lepidozygus tapeinosoma</i>	Planktivore
<i>Neoglyphidodon bonang</i>	Planktivore & Herbivore
<i>Neoglyphidodon carlsoni</i>	Planktivore & Herbivore
<i>Neoglyphidodon melas</i>	Planktivore & Herbivore
<i>Neoglyphidodon nigroris</i>	Planktivore & Herbivore
<i>Neoglyphidodon polyacanthus</i>	Planktivore & Herbivore
<i>Neoglyphidodon sp.</i>	Planktivore & Herbivore
<i>Neoglyphidodon thoracotaeniatus</i>	Planktivore & Herbivore
<i>Neopomacentrus azysron</i>	Planktivore
<i>Neopomacentrus bankieri</i>	Planktivore
<i>Neopomacentrus cyanomos</i>	Planktivore
<i>Neopomacentrus sp.</i>	Planktivore
<i>Neopomacentrus violascens</i>	Planktivore
<i>Plectroglyphidodon dickii</i>	Omnivore
<i>Plectroglyphidodon lacrymatus</i>	Omnivore
<i>Plectroglyphidodon leucozonus</i>	Omnivore
<i>Plectroglyphidodon phoenixensis</i>	Herbivore
<i>Plectroglyphidodon plectroglyphidodon</i>	Herbivore
<i>Pomacentridae brachialis</i>	Planktivore & Herbivore
<i>Pomacentridae imitator</i>	Planktivore & Herbivore
<i>Pomacentrus adelus</i>	Planktivore & Herbivore
<i>Pomacentrus albicaudatus</i>	Planktivore & Herbivore
<i>Pomacentrus amboinensis</i>	Planktivore & Herbivore
<i>Pomacentrus azyrson</i>	Planktivore & Herbivore
<i>Pomacentrus bankanensis</i>	Planktivore & Herbivore
<i>Pomacentrus brachialis</i>	Planktivore & Herbivore
<i>Pomacentrus chrysurus</i>	Planktivore & Herbivore
<i>Pomacentrus coelestis</i>	Planktivore & Herbivore
<i>Pomacentrus colini</i>	Planktivore & Herbivore
<i>Pomacentrus cuneatus</i>	Planktivore & Herbivore
<i>Pomacentrus lepidogenys</i>	Planktivore & Herbivore
<i>Pomacentrus littoralis</i>	Planktivore & Herbivore
<i>Pomacentrus milleri</i>	Planktivore & Herbivore
<i>Pomacentrus moluccensis</i>	Planktivore & Herbivore
<i>Pomacentrus nigromanus</i>	Planktivore & Herbivore
<i>Pomacentrus pavo</i>	Planktivore & Herbivore
<i>Pomacentrus philippinus</i>	Planktivore & Herbivore
<i>Pomacentrus polyspines</i>	Planktivore & Herbivore
<i>Pomacentrus proteus</i>	Planktivore & Herbivore
<i>Pomacentrus similis</i>	Planktivore & Herbivore
<i>Pomacentrus simsiang</i>	Planktivore & Herbivore

Pomacentrus sp.	Planktivore & Herbivore
Pomacentrus spilotoceps	Planktivore & Herbivore
Pomacentrus trichourus	Planktivore & Herbivore
Pomacentrus tripunctatus	Planktivore & Herbivore
Pomacentrus vaiula	Planktivore & Herbivore
Pomacentrus violascens	Planktivore & Herbivore
Pomachromis richardsoni	Planktivore & Herbivore
Premnas biaculeatus	Planktivore & Herbivore
Stegastes albifasciatus	Herbivore
Stegastes fasciolatus	Herbivore
Stegastes gascoynei	Herbivore
Stegastes lividus	Herbivore
Stegastes nigricans	Herbivore
Stegastes obreptus	Herbivore
<b>Priacanthidae</b>	
Priacanthus hamrur	Invertebrate feeder & Piscivore
<b>Scaridae</b>	
Calotomus carolinus	Herbivorous
Calotomus spinidens	Herbivorous
Cetoscarus bicolor	Herbivorous
Chlorurus bleekeri	Herbivorous
Chlorurus bowersi	Herbivorous
Chlorurus capistratoides	Herbivorous
Chlorurus gibbus	Herbivorous
Chlorurus japonensis	Herbivorous
Chlorurus microrhinos	Herbivorous
Chlorurus sordidus	Herbivorous
Chlorurus sp.	Herbivorous
Chlorurus troschellii	Herbivorous
Hipposcarus harid	Herbivorous
Hipposcarus longiceps	Herbivorous
Leptoscarus vaigiensis	Herbivorous
Scarus niger	Herbivorous
Scarus altipinnis	Herbivorous
Scarus bleekeri	Herbivorous
Scarus chameleon	Herbivorous
Scarus dimidiatus	Herbivorous
Scarus flavipectoralis	Herbivorous
Scarus forsteni	Herbivorous
Scarus frenatus	Herbivorous
Scarus ghobban	Herbivorous
Scarus globiceps	Herbivorous
Scarus niger	Herbivorous
Scarus oviceps	Herbivorous

Scarus prasiognathos	Herbivorous
Scarus psittacus	Herbivorous
Scarus quoyi	Herbivorous
Scarus rivulatus	Herbivorous
Scarus rubroviolaceus	Herbivorous
Scarus russelii	Herbivorous
Scarus schlegeli	Herbivorous
Scarus spinus	Herbivorous
Scarus stronglycephalus	Herbivorous
Scarus tricolor	Herbivorous
Scarus xanthopleura	Herbivorous
<b>Scombridae</b>	
Rastrelliger kanagurta	Planktivore
Sarda orientalis	Invertebrate & Piscivore
Scomberomorus commerson	Piscivore
Thunnus obesus	Invertebrate feeder & Piscivore
<b>Scorpaenidae</b>	
Pterois volitan	Planktivore & Invertebrate
<b>Serranidae</b>	
Anyperodon leucogrammicus	Invertebrate feeder & Piscivore
Cephalopholis argus	Piscivore
Cephalopholis boenack	Invertebrate feeder & Piscivore
Cephalopholis urodeta	Invertebrate feeder & Piscivore
Epinephelus areolatus	Invertebrate feeder & Piscivore
Epinephelus bontoides	Invertebrate feeder & Piscivore
Epinephelus caeruleopunctatus	Invertebrate feeder & Piscivore
Epinephelus coiodes	Invertebrate feeder & Piscivore
Epinephelus longispinis	Invertebrate feeder & Piscivore
Epinephelus maculatus	Invertebrate feeder & Piscivore
Epinephelus malabaricus	Invertebrate feeder & Piscivore
Epinephelus merra	Invertebrate feeder & Piscivore
Epinephelus polyphkadion	Invertebrate feeder & Piscivore
Epinephelus sp	Invertebrate feeder & Piscivore
Gracila albormarginata	Invertebrate feeder
Plectropomus areolatus	Piscivore
Plectropomus leopardus	Invertebrate feeder & Piscivore
Plectropomus maculatus	Invertebrate feeder & Piscivore
Plectropomus pessuliferus	Piscivore
Pseudanthias pascalus	Planktivore
Variola louti	Invertebrate feeder & Piscivore
<b>Siganidae</b>	
Siganus argenteus	Herbivorous
Siganus corallinus	Herbivorous
Siganus doliatus	Herbivorous

<i>Siganus guttatus</i>	Herbivorous
<i>Siganus puelloides</i>	Omnivore
<i>Siganus punctatissimus</i>	Herbivorous
<i>Siganus punctatus</i>	Herbivorous
<i>Siganus sp.</i>	Herbivorous
<i>Siganus spinus</i>	Herbivorous
<i>Siganus stellatus</i>	Herbivorous
<i>Siganus uspi</i>	Herbivorous
<i>Siganus vermiculatus</i>	Herbivorous
<i>Siganus virgatus</i>	Herbivorous
<i>Siganus vulpinus</i>	Herbivorous
<b>Soleidae</b>	
<i>Soleidae sp.</i>	Invertebrate feeder
<b>Sparidae</b>	
<i>Sarpa salpa</i>	Omnivore
<b>Sphyraenidae</b>	
<i>Sphyraena qenie</i>	Invertebrate feeder & Piscivore
<i>Sphyraena sp.</i>	Invertebrate feeder & Piscivore
<b>Syngnathidae</b>	
<i>Corythoichthys haematopterus</i>	Planktivore
<b>Synodontidae</b>	
<i>Saurida nebulosa</i>	Piscivore
<i>Synodus dermatogenys</i>	Invertebrate feeder
<i>Synodus variegatus</i>	Invertebrate feeder
<b>Terapontidae</b>	
<i>Terapon jarbua</i>	Omnivore
<b>Tetraodontidae</b>	
<i>Arothron hispidus</i>	
<i>Arothron meleagris</i>	Corrallivore & Invertebrate
<i>Arothron nigropunctatus</i>	Corrallivore & Invertebrate
<i>Arothron sp.</i>	Corrallivore & Invertebrate
<i>Canthigaster amboinensis</i>	Omnivore
<i>Canthigaster bennetti</i>	Omnivore
<i>Canthigaster coronata</i>	Omnivore
<i>Canthigaster ocellicincta</i>	Omnivore
<i>Canthigaster solandri</i>	Omnivore
<i>Canthigaster valentini</i>	Omnivore
<b>Zanclidae</b>	
<i>Zanclus cornutus</i>	Invertebrate feeder